

APPLIED COST-EFFECTIVENESS MODELING WITH R

SMDM

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Learning objectives

- Understand how R can be used to perform model-based cost-effectiveness analysis with existing packages;
- Develop own models in R by modifying existing code for commonly used model types;
- Understand how using R can improve reproducibility and transparency of model-based cost-effectiveness analysis

Agenda

- Introduction
- Basic model taxonomy
- Simple Markov cohort model
- Semi-Markov multi-state model
- Partitioned-survival model
- Cost-effectiveness analysis
- Summary

Structure

- Presentation
- Exercises using R

Online tutorial

rcea 0.1.2

Reference

Tutorials ▾

Slides



rcea

This is the repository for the `rcea` package. A wide range of models are covered in this repository, including individual patient data (IPD) and semi-Markov individual patient data (IPD) models. Sensitivity analysis can be used to evaluate the robustness of the results.

Simple Markov Cohort Model

Incorporating Probabilistic Sensitivity Analysis

Markov Cohort Model with hesim

Semi-Markov Multi-state Model

Partitioned Survival Model

Cost-effectiveness Analysis

Model-based cost-effectiveness analysis (CEA) with R. A range of models are covered in this repository, including individual patient data (IPD) and semi-Markov individual patient data (IPD) models. Sensitivity analysis can be used to evaluate the robustness of the results.

The course materials are available at <https://hesim-dev.github.io/rcea>.

Installation and setup

All required R packages and course materials can be installed with the following steps.

1. Open an R session. We recommend using [RStudio](#).
2. Install the `rcea` package from GitHub, which will also install all other required packages.

```
# install.packages("devtools") # You must install the "devtools" R package first.
devtools::install_github("hesim-dev/rcea")
```

3. Create a new project in your desired directory.

```
# Create a project named "rcea-exercises" within a directory named "Projects"
usethis::create_project("~/Projects/rcea-exercises")
```

4. Add the course materials (R scripts for the tutorials) to your new project.

```
rcea::use_rcea("~/Projects/rcea-exercises")
```

Links

Browse source code at

<https://github.com/hesim-dev/rcea/>

Report a bug at

<https://github.com/hesim-dev/rcea/issues>

License

[GPL-3](#)

Developers

Devin Incerti

Author, maintainer

Jeroen P. Jansen

Author

R scripts for exercises

The screenshot shows a Mac OS X Finder window titled "Projects". The sidebar on the left lists "Favorites" including Google, Box, Macintosh HD, Applications, Desktop, Downloads, and Jero... (with a triangle icon). The main pane displays a file list with the following structure:

Name	Date Modified	Size	Kind
▼ rcea-exercises	Today at 4:30 PM	--	Folder
01-markov-cohort.R	Today at 4:30 PM	4 KB	R Source File
02-markov-cohort-psa.R	Today at 4:30 PM	6 KB	R Source File
03-markov-cohort-hesim.R	Today at 4:30 PM	4 KB	R Source File
04-mstate.R	Today at 4:30 PM	5 KB	R Source File
05-psm.R	Today at 4:30 PM	4 KB	R Source File
06-cea.R	Today at 4:30 PM	5 KB	R Source File
► R	Today at 4:29 PM	--	Folder
rcea-exercises.Rproj	Today at 4:29 PM	276 bytes	R Project

RStudio Cloud

Your Workspace / RCEA

RAM    Jeroen Jansen

File Edit Code View Plots Session Build Debug Profile Tools Help

   Go to file/function

R 4.1.0

01-markov-cohort.R x 02-markov-cohort-psa.R x 03-markov-cohort-hesim.R x 04-mstate.R x 05-psm.R x 06-cea.R x

Run Source

```
1 ## ---- Overview
2 ## @knitr R-packages
3 library("rcea")
4 library("knitr")
5 library("kableExtra")
6 library("magrittr")
7 library("tibble")
8
9 ## ---- Model parameters -----
10 ## @knitr transition-probabilities
11 p_hd <- .002 # constant probability of dying when Healthy (all-cause mortality)
12 p_hs1 <- .15 # probability of becoming Sick when Healthy
13 p_s1h <- .05 # probability of becoming Healthy when Sick
14 p_s1s2 <- .105 # probability of becoming Sicker when Sick
15 p_s1d <- .006 # constant probability of dying when Sick
16 p_s2d <- .02 # constant probability of dying when Sicker
17
18 ## @knitr transition-probability-complements
19 p_hh <- 1 - p_hs1 - p_hd
20 p_s1s1 <- 1 - p_s1h - p_s1s2 - p_s1d
21 p_s2s2 <- 1 - p_s2d
```

1:1  Overview

Environment History Connections Tutorial

  293 MiB  List 

R Global Environment 

Environment is empty

Files Plots Packages Help Viewer

 New Folder  Upload  Delete  Rename

 Home > Projects > rcea-exercises 

Name	Size
 ..	
 .gitignore	12 B
 R	
 rcea-exercises.Rproj	276 B
 01-markov-cohort.R	4.1 KB
 02-markov-cohort-psa.R	5.5 KB
 03-markov-cohort-hesim.R	3.4 KB
 04-mstate.R	4.5 KB
 05-psm.R	4 KB
 06-cea.R	4.7 KB

Console Terminal x Jobs x

R 4.1.0 · ~/Projects/rcea-exercises/ 

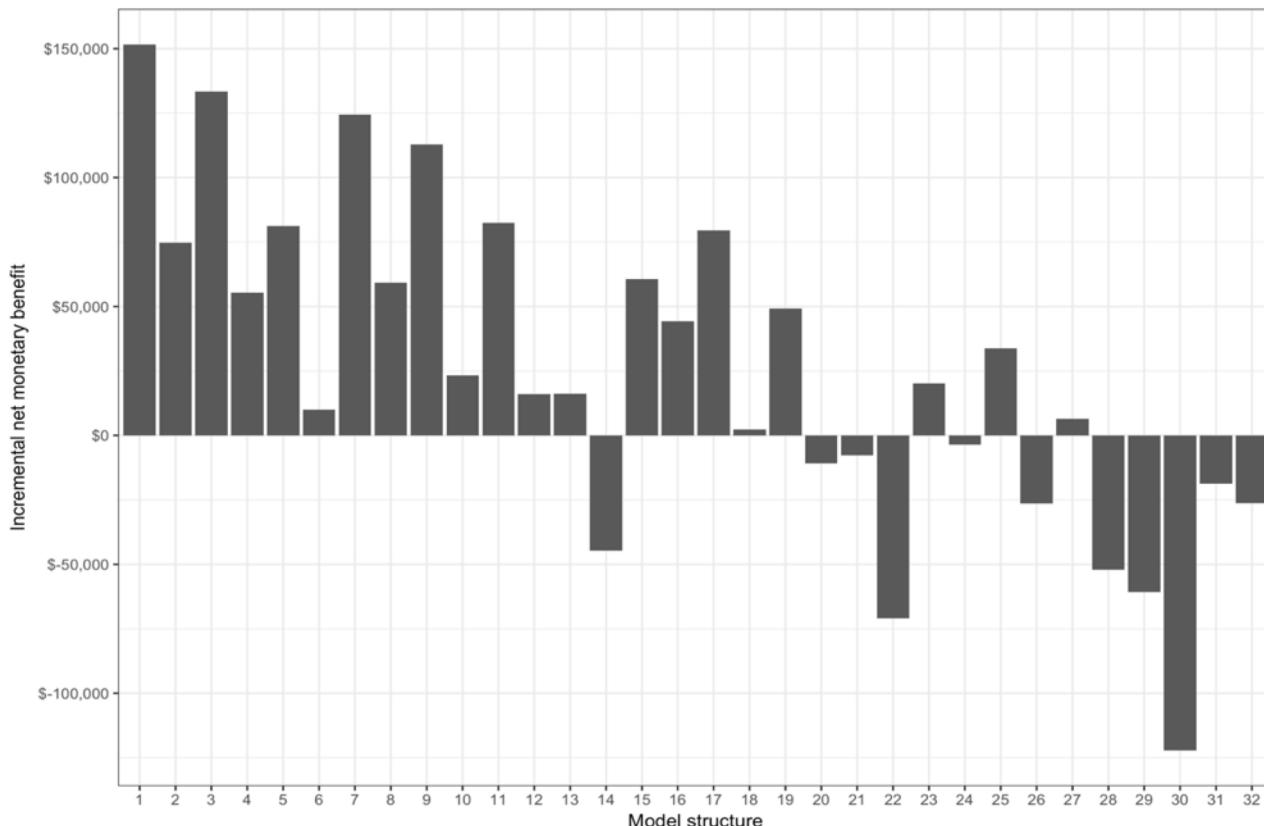
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Introduction

Criteria that economic models should strive to meet

- Clinical realism
 - A model should reflect the state of evidence, the current understanding of the disease, and be accepted by clinical experts.
- Quantifying decision uncertainty
 - A model should be capable of quantifying decision uncertainty and informing prioritization of future research.
- Transparency and reproducibility
 - Resources should exist so that a model can be completely understood, reproduced, and pressure tested.
- Reusability and adaptability
 - It should be possible to easily update a model to reflect new clinical evidence or adapt it for a new market, indication, or intervention.

Structural uncertainty

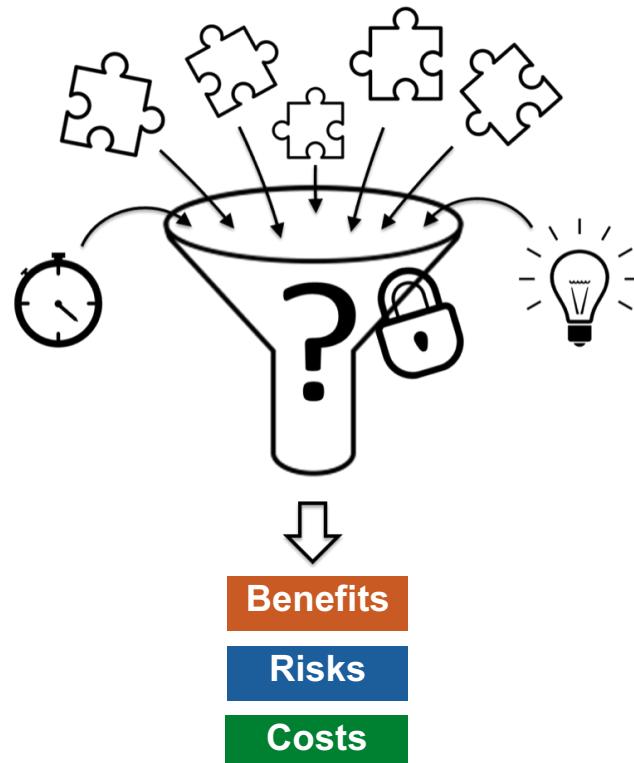


Plugging in model input parameter estimates



What do we mean with model transparency?

- Concept, math
- Face validity
- Implementation/programming
- Open-source, open-access
- *Familiarity with software?*



Modeling in Excel

- Excel has been dominant software platform used by HE modelers, especially for HTA submissions
- Reasons are not surprising
 - Practically everyone with a computer has access to Excel
 - Does not require that you learn a new programming language
- Many consider its “transparency” to be an attribute
- With models in Excel, you can follow calculations that are being performed in every single cell of every single worksheet

The toolbar includes icons for Paste, Cut, Copy, Format, Arial font, font size 9, bold, italic, underline, alignment options, Wrap Text, Number, Conditional Formatting, Format as Table, Cell Styles, Insert, Delete, Format, AutoSum, Fill, Clear, and Sort & Filter.

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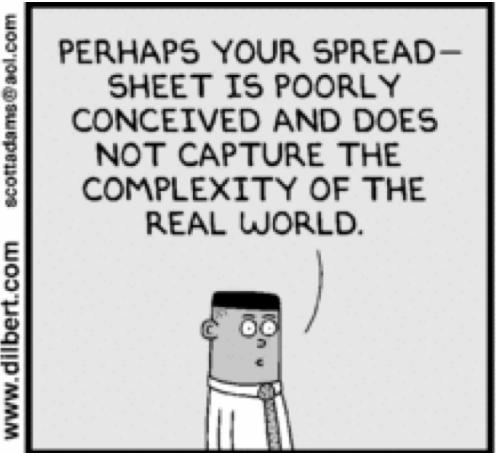
A	B	C	D	E	F	G	H	I	J	K	L	M	N	O	P	Q	R	S	T	U	V
PSA INPUTS																					
Efficacy and transitions between health states (other than death)		point estimate	S(D)estimate)		low	high	distribution	alpha	beta	Source	sampled value from uncertainty distribution	value 'plugged in' model			One way sensitivity input low	One way sensitivity input high					
PROBABILITY Induction response Standard non-biologic treatment	0.355	0.017		0.322	0.390	beta	267.30	484.82			0.3562	0.3554			0.32	0.39					
OR Induction response Adalimumab 160/80/40 vs Standard non-biologic treatment	1.870		0.957		1.654	log-normal					1.9728	1.8700			0.96	3.65					
OR Induction response Golimumab 200/100/50(100) vs Standard non-biologic treatment	2.115		1.011		3.953	log-normal					1.9268	2.1150			1.01	3.95					
OR Induction response Infliximab 5mg/kg vs Standard non-biologic treatment	4.118		2.084		8.144	log-normal					2.4537	4.1180			2.08	8.14					
PROBABILITY Induction remission Standard non-biologic treatment	0.089	0.010	0.071		0.110	beta	71.65	729.91			0.0870	0.0894			0.07	0.11					
OR Induction remission Adalimumab 160/80/40 vs Standard non-biologic treatment	2.254		1.076		4.717	log-normal					2.8307	2.2540			1.08	4.72					
OR Induction remission Golimumab 200/100/50(100) vs Standard non-biologic treatment	3.998		1.324		6.277	log-normal					3.5223	2.9890			1.32	6.28					
OR Induction remission Infliximab 5mg/kg vs Standard non-biologic treatment	5.271		2.596		11.640	log-normal					5.2710	5.2710			2.60	11.64					
PROBABILITY sustained response Standard non-biologic treatment	0.829	0.016	0.797		0.858	beta	489.00	101.18			0.8459	0.8286			0.80	0.86					
OR sustained response Adalimumab 160/80/40	1.311		0.669		2.390	log-normal					1.9203	1.3110			0.67	2.59					
OR sustained response Golimumab 200/100/50(100)	1.623		1.067		2.302	log-normal					1.9808	1.6230			1.07	2.50					
OR sustained response Infliximab 5mg/kg	2.116		1.021		4.540	log-normal					2.8782	2.1160			1.02	4.54					
PROBABILITY sustained remission Standard non-biologic treatment	0.861	0.026	0.806		0.908	beta	151.60	24.49			0.9124	0.8669			0.81	0.91					
OR sustained remission Adalimumab 160/80/40	0.762		0.217		2.556	log-normal					0.3871	0.7624			0.22	2.56					
OR sustained remission Golimumab 200/100/50(100)	0.918		0.359		2.452	log-normal					1.1049	0.9180			0.36	2.45					
OR sustained remission Infliximab 5mg/kg	1.300		0.435		4.053	log-normal					1.7175	1.3000			0.44	4.05					
Proportion transition to relapse when losing remission status	0.000	0.000	0NUM!	0NUM!	0NUM!	beta	0.00	-1.00			0.0000	0.0000			0.00	0.00					
PROBABILITY maintenance discontinuation Standard non-biologic treatment	0.000	0.000	0NUM!	0NUM!	0NUM!	beta	0.00	-1.00			0.0000	0.0000			0.00	0.00					
OR maintenance discontinuation Adalimumab 160/80/40	0.000		0.000		0.000	log-normal					0NUM!	0.0000			0.00	0.00					
OR maintenance discontinuation Golimumab 200/100/50(100)	0.000		0.000		0.000	log-normal					0NUM!	0.0000			0.00	0.00					
OR maintenance discontinuation Infliximab 5mg/kg	0.000		0.000		0.000	log-normal					0NUM!	0.0000			0.00	0.00					
PROBABILITY success 1 round IV steroids	0.480	0.007	0.468		0.493	beta	2767.63	2993.46			0.4859	0.4804			0.47	0.49					
PROBABILITY success with 2nd round IV steroids given failure 1st round	0.480	0.007	0.468		0.493	beta	2767.63	2993.46			0.4799	0.4804			0.47	0.49					
PROBABILITY loss of response (relapse management)	0.171	0.016	0.142		0.203	beta	101.18	489.00			0.1751	0.1714			0.14	0.20					
PROBABILITY long term complications given remission (post-colectomy)	0.015	0.007	0.004		0.032	beta	4.41	290.51			0.0056	0.0149			0.00	0.03					
PROBABILITY successful recovery from long-term complications (post-colectomy)	1.000	0.000	0NUM!	0NUM!	0NUM!	beta	-1.00	0.00			1.0000	1.0000			1.00	1.00					
Safety (induction treatment)																					
PROBABILITY serious infections Standard non-biologic treatment	0.016	0.003	0.011		0.021	beta	36.84	2283.37			0.0161	0.0159			0.01	0.02					
OR serious infections Adalimumab 160/80/40 vs Standard non-biologic treatment	1.100		0.830		1.460	log-normal					1.2132	1.1000			0.83	1.46					
OR serious infections Golimumab 200/100/50(100) vs Standard non-biologic treatment	1.100		0.830		1.460	log-normal					1.0408	1.1000			0.83	1.46					
OR serious infections Infliximab 5mg/kg vs Standard non-biologic treatment	1.100		0.830		1.460	log-normal					1.2286	1.1000			0.83	1.46					
PROBABILITY Adverse event 2 Standard non-biologic treatment	0.000	0.000	0NUM!	0NUM!	0NUM!	beta	0.00	-1.00			0.0000	0.0000			0.00	0.00					
OR Adverse event 2 Adalimumab 160/80/40 vs Standard non-biologic treatment	1.000		1.000		1.000	log-normal					1.0000	1.0000			1.00	1.00					
OR Adverse event 2 Golimumab 200/100/50(100) vs Standard non-biologic treatment	1.000		1.000		1.000	log-normal					1.0000	1.0000			1.00	1.00					
OR Adverse event 2 Infliximab 5mg/kg vs Standard non-biologic treatment	1.000		1.000		1.000	log-normal					1.0000	1.0000			1.00	1.00					
PROBABILITY Adverse event 3 Standard non-biologic treatment	0.000	0.000	0NUM!	0NUM!	0NUM!	beta	0.00	-1.00			0.0000	0.0000			0.00	0.00					
OR Adverse event 3 Adalimumab 160/80/40 vs Standard non-biologic treatment	1.000		1.000		1.000	log-normal					1.0000	1.0000			1.00	1.00					
OR Adverse event 3 Golimumab 200/100/50(100) vs Standard non-biologic treatment	1.000		1.000		1.000	log-normal					1.0000	1.0000			1.00	1.00					
OR Adverse event 3 Infliximab 5mg/kg vs Standard non-biologic treatment	1.000		1.000		1.000	log-normal					1.0000	1.0000			1.00	1.00					
PROBABILITY Adverse event 4 Standard non-biologic treatment	0.000	0.000	0NUM!	0NUM!	0NUM!	beta	0.00	-1.00			0.0000	0.0000			0.00	0.00					
OR Adverse event 4 Adalimumab 160/80/40 vs Standard non-biologic treatment	1.000		1.000		1.000	log-normal					1.0000	1.0000			1.00	1.00					
OR Adverse event 4 Golimumab 200/100/50(100) vs Standard non-biologic treatment	1.000		1.000		1.000	log-normal					1.0000	1.0000			1.00	1.00					
OR Adverse event 4 Infliximab 5mg/kg vs Standard non-biologic treatment	1.000		1.000		1.000	log-normal					1.0000	1.0000			1.00	1.00					

The screenshot shows the Microsoft Excel ribbon with the 'Home' tab selected. The ribbon includes sections for 'Clipboard' (Cut, Copy, Paste, Format), 'Font' (Font, Size, Bold, Italic, Underline, Font Color, Font Style, Alignment, Number Format, Wrap Text, Merge & Center), 'Cells' (Conditional Formatting, Format as Table, Cell Styles, Insert, Delete, Format), and 'Styles' (AutoSum, Fill, Clear, Sort & Filter).

Where is Waldo



Time to change?



Alternative



BCEA

heemod

hesim

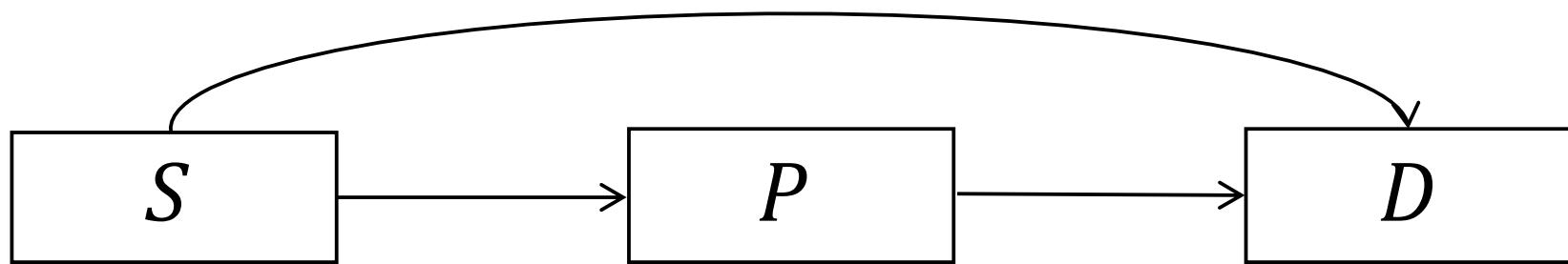
...

What is R

- Statistical programming language and environment for statistical computing
- Free to use (open source, user developed packages that are transparent)
- Very good for data management, statistical analysis, and visualization
- Scripts contain all steps to perform an analysis
- CEA models can be coded from ‘scratch’ using base R or via convenient and improving packages

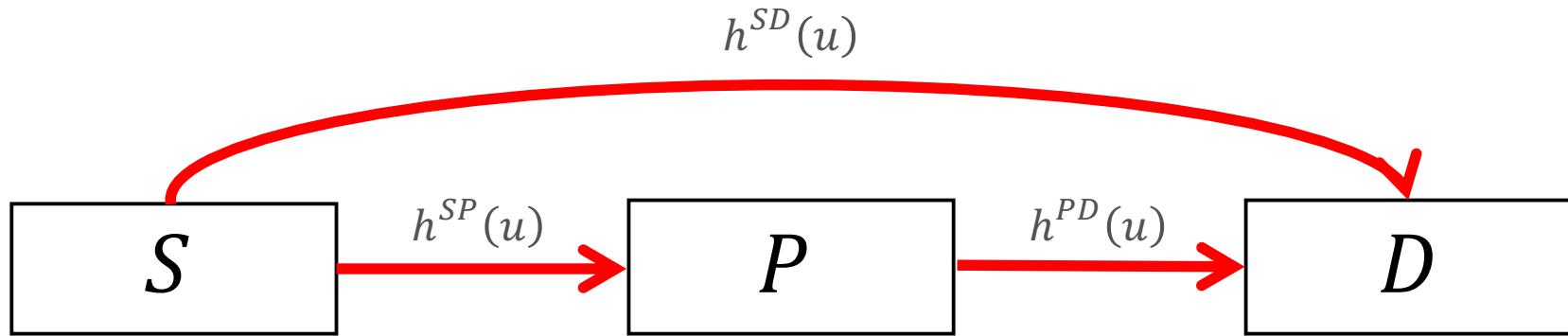
Basic model taxonomy

Health states describing course of disease over time



Markov model

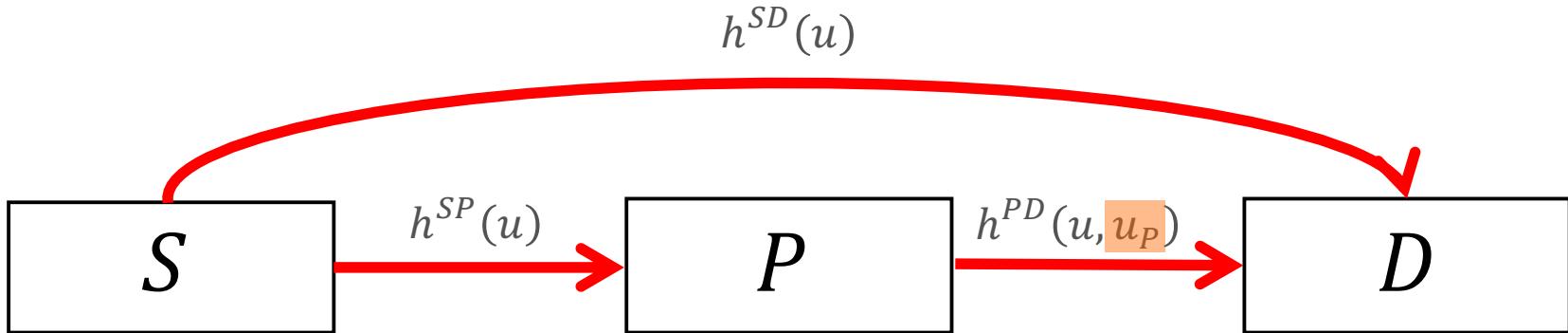
Clock forward



transition rates depend only on time in model

Semi-Markov model

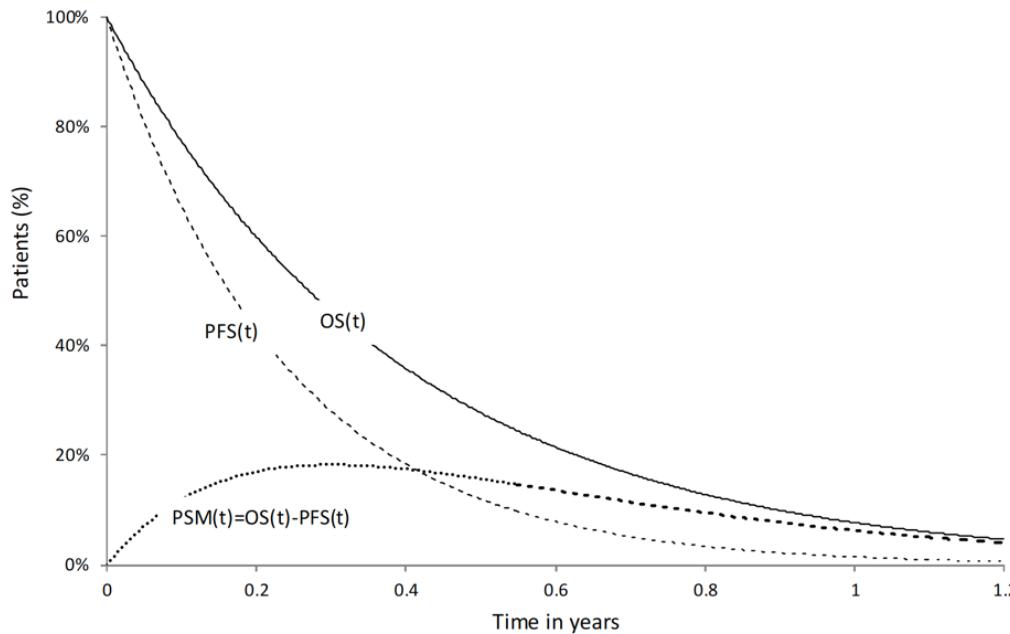
Clock reset



transition rates depend on time in model

some transitions depend on time in an intermediate health state

Partitioned survival model



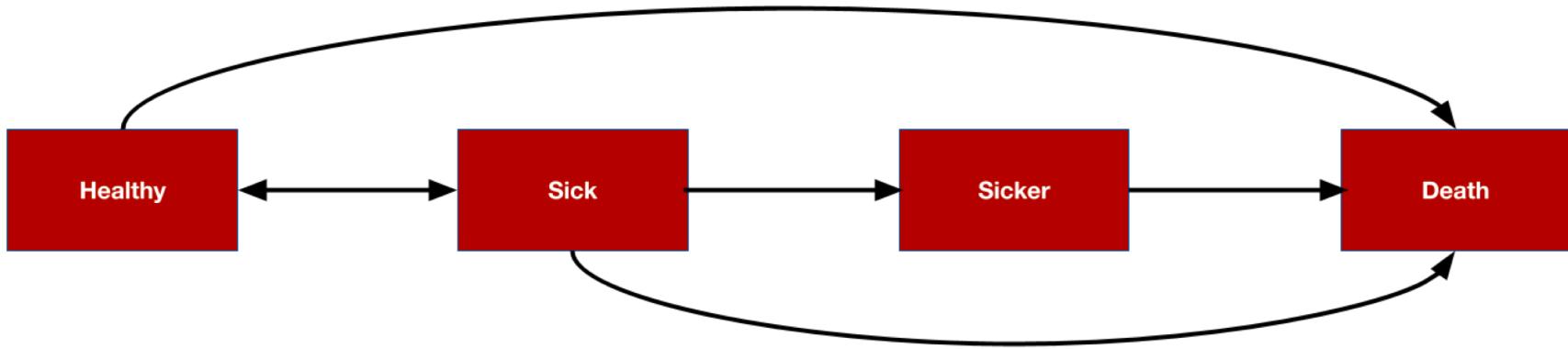
Summary of model types

			discrete time	continuous time
State transition models	Markov ("Clock forward")	Cohort	cohort discrete time state transition models (cDTSTM) <ul style="list-style-type: none"> • time-homogeneous Markov models • time-inhomogeneous Markov models 	cohort continuous time state transition models
		Individual-level	individual-level discrete time state transition models (iDTSTM)	individual-level continuous time state transition models (iCTSTM)
	Semi-Markov ("Clock reset")*	Individual-level	iDTSTM	iCTSTM
Partitioned survival model		Cohort		✓

*tunnel states can be used in a cohort model to approximate a semi-Markov process

Simple Markov cohort model

Model



Model

- Annual transition probabilities with SOC

	Healthy	Sick	Sicker	Death
Healthy	0.848	0.15	0	0.002
Sick	0.5	0.389	0.105	0.006
Sicker	0	0	0.98	0.02
Death	0	0	0	1.000

- Relative risk of progression to a worse health state with new intervention is 0.8

- Drug costs

	New	SOC
Drug costs	12000	2000

- Other annual costs and utility

	Healthy	Sick	Sicker	Death
Direct medical	2000	4000	15000	0
Utility	1	0.75	0.5	0

- Annual discount rates of 3% for costs and 3% for QALYs

SOC

	Healthy	Sick	Sicker	Death	Cycle	Healthy	Sick	Sicker	Death	discounted QALYs	discounted medical costs	discounted treatment costs	discounted total costs
Healthy	0.848	0.15	0	0.002	0	1.00000	0.00000	0.00000	0.00000	1.00000	2000.000	2000	
Sick	0.5	0.389	0.105	0.006	1	0.84800	0.15000	0.00000	0.00200	0.93252	2229.126	1937.864	
Sicker	0	0	0.98	0.02	2	0.79410	0.18555	0.01575	0.00460	0.88712	2419.321	1876.527	
Death	0	0	0	1	3	0.76618	0.19129	0.03492	0.00761	0.84843	2581.884	1816.350	
					4	0.74536	0.18934	0.05431	0.01099	0.81254	2721.140	1757.443	
					5	0.72674	0.18546	0.07310	0.01470	0.77840	2839.541	1699.850	
					6	0.70900	0.18115	0.09111	0.01873	0.74572	2938.972	1643.593	
	Healthy	Sick	Sicker	Death	80	0.11578	0.02960	0.29714	0.55748	0.02693	451.752	83.173	
Utility	1	0.75	0.5	0	81	0.11298	0.02888	0.29430	0.56384	0.02571	433.941	79.591	
Direct medical	2000	4000	15000	0	82	0.11025	0.02818	0.29145	0.57012	0.02455	416.778	76.159	
					83	0.10758	0.02750	0.28858	0.57634	0.02344	400.244	72.871	
Drug costs	2000				84	0.10498	0.02683	0.28570	0.58249	0.02237	384.317	69.722	
					85	0.10244	0.02619	0.28280	0.58858	0.02136	368.979	66.704	
					Sum					21.08			204,123

Steps

- Define transition matrix SOC
- Define transition matrix New treatment
- Define utility and cost values by health state
- Calculate health state probabilities over time
- Calculate expected QALYs and costs
- Cost-effectiveness analysis

Define transition matrix with standard of care

```
p_hd <- 0.002          # constant probability of dying when Healthy (all-cause mortality)
p_hs1 <- 0.15           # probability of becoming Sick when Healthy
p_s1h <- 0.5             # probability of becoming Healthy when Sick
p_s1s2 <- 0.105          # probability of becoming Sicker when Sick
p_s1d <- 0.006          # constant probability of dying when Sick
p_s2d <- 0.02            # constant probability of dying when Sicker

p_hh <- 1 - p_hs1 - p_hd
p_s1s1 <- 1 - p_s1h - p_s1s2 - p_s1d
p_s2s2 <- 1 - p_s2d
```

```
p_soc <- matrix(
  c(p_hh,  p_hs1,  0,      p_hd,
    p_s1h,  p_s1s1,  p_s1s2, p_s1d,
    0,      0,      p_s2s2, p_s2d,
    0,      0,      0,      1),
  byrow = TRUE,
  nrow = 4, ncol = 4
)
state_names <- c("H", "S1", "S2", "D")
colnames(p_soc) <- rownames(p_soc) <- state_names

print(p_soc)
```

	H	S1	S2	D
H	0.848	0.150	0.000	0.002
S1	0.500	0.389	0.105	0.006
S2	0.000	0.000	0.980	0.020
D	0.000	0.000	0.000	1.000

Relative risk and transition matrix with New treatment

```
apply_rr <- function(p, rr = .8){  
  p["H", "S1"] <- p["H", "S1"] * rr  
  p["H", "S2"] <- p["H", "S2"] * rr  
  p["H", "D"] <- p["H", "D"] * rr  
  p["H", "H"] <- 1 - sum(p["H", -1])  
  
  p["S1", "S2"] <- p["S1", "S2"] * rr  
  p["S1", "D"] <- p["S1", "D"] * rr  
  p["S1", "S1"] <- 1 - sum(p["S1", -2])  
  
  p["S2", "D"] <- p["S2", "D"] * rr  
  p["S2", "S2"] <- 1 - sum(p["S2", -3])  
  
  return(p)  
}  
  
p_new <- apply_rr(p_soc, rr = .8)
```

	H	S1	S2	D
H	0.8784	0.1200	0.000	0.0016
S1	0.5000	0.4112	0.084	0.0048
S2	0.0000	0.0000	0.984	0.0160
D	0.0000	0.0000	0.000	1.0000

Utility and costs

```
utility <- c(1, .75, .5, 0)
costs_medical <- c(2000, 4000, 15000, 0)
costs_treat_soc <- c(rep(2000, 3), 0)
costs_treat_new <- c(rep(12000, 3), 0)
```

```
> utility
[1] 1.000 0.75 0.500 0.000
```

```
> costs_medical
[1] 2000 4000 15000 0
```

```
> costs_treat_soc
[1] 2000 2000 2000 0
```

```
> costs_treat_new
[1] 12000 12000 12000 0
```

Simulation – health state probabilities

Matrix multiplication

```
x_init <- c(1, 0, 0, 0)  
x_init %*% p_soc
```

```
H      S1     S2      D  
[1,] 0.848  0.15    0  0.002
```

```
x_init %*% p_soc %*% p_soc
```

```
H      S1      S2      D  
[1,] 0.794104 0.18555 0.01575 0.004596
```

Simulation – health state probabilities with a function

x0 The state vector at model cycle 0 (i.e., the initial state vector)
p The transition probability matrix
n_cycles The number of model cycles. (Default is 85)

```
sim_markov_chain <- function(x0, p, n_cycles = 85){

  x <- matrix(NA, ncol = length(x0), nrow = n_cycles) # Initialize Markov trace
  x <- rbind(x0, x)                                     # Markov trace at cycle 0 is initial state vector
  colnames(x) <- colnames(p)                            # Columns are the health states
  rownames(x) <- 0:n_cycles                             # Rows are the model cycles

  for (t in 1:n_cycles){                                # Simulating state vectors at each cycle with for loop
    x[t + 1, ] <- x[t, ] %*% p
  }

  return(x)
}
```

Simulation – health state probabilities with a function

```
x_soc <- sim_markov_chain(x_init, p_soc)
```

	H	S1	S2	D
0	1.0000000	0.0000000	0.0000000	0.000000000
1	0.8480000	0.1500000	0.0000000	0.002000000
2	0.7941040	0.18555000	0.01575000	0.004596000
3	0.7661752	0.19129455	0.03491775	0.007612508
4	0.7453638	0.18933986	0.05430532	0.010990981
5	0.7267385	0.18545778	0.07309990	0.014703854
6	0.7090031	0.18115385	0.09111097	0.018732076
7	0.6918116	0.17681931	0.10830990	0.023059224
8	0.6750659	0.17255445	0.12470973	0.027669961
9	0.6587331	0.16838356	0.14033376	0.032549614

79	0.1186504	0.03032935	0.29995426	0.551065992
80	0.1157802	0.02959568	0.29713976	0.557484354
81	0.1129795	0.02887975	0.29430451	0.563836283
82	0.1102465	0.02818114	0.29145079	0.570121611
83	0.1075796	0.02749943	0.28858079	0.576340207
84	0.1049772	0.02683421	0.28569662	0.582491978
85	0.1024378	0.02618509	0.28280028	0.588576870

```
x_new <- sim_markov_chain(x_init, p_new)
```

	H	S1	S2	D
0	1.0000000	0.0000000	0.0000000	0.000000000
1	0.8784000	0.1200000	0.0000000	0.001600000
2	0.8315866	0.15475200	0.01008000	0.003581440
3	0.8078416	0.16342441	0.02291789	0.005816068
4	0.7913203	0.16414111	0.03627885	0.008259738
5	0.7771663	0.16245326	0.04948624	0.010894190
6	0.7638895	0.16006074	0.06234054	0.013709211
7	0.7510309	0.15748372	0.07478819	0.016697175
8	0.7384474	0.15488101	0.08682021	0.019851357
9	0.7260927	0.15230076	0.09844109	0.023165425

79	0.2230518	0.04678750	0.31880122	0.411359506
80	0.2193224	0.04600523	0.31763055	0.417041788
81	0.2156554	0.04523604	0.31641290	0.422695618
82	0.2120498	0.04447971	0.31515012	0.428320406
83	0.2085044	0.04373603	0.31384402	0.433915590
84	0.2050182	0.04300478	0.31249634	0.439480634
85	0.2015904	0.04228575	0.31110880	0.445015028

Computing a present value with a function

```
pv <- function(z, dr, t) {  
  z/(1 + dr)^t  
}  
  
z   A numeric quantity  
dr  Discount rate  
t   Vector of times to compute the present value
```

```
pv(1000, dr = .03, t = 0:4)
```

```
[1] 1000.0000 970.8738 942.5959 915.1417 888.4870
```

QALYs after 1st cycle

```
x_soc[2, ] # State occupancy probabilities after 1st cycle
```

H	S1	S2	D
0.848	0.150	0.000	0.002

```
sum(x_soc[2, 1:3]) # Expected life-years after 1st cycle
```

```
[1] 0.998
```

```
sum(x_soc[2, ] * utility) # Expected utility after 1st cycle
```

```
[1] 0.9605
```

```
sum(pv(x_soc[2, ] * utility, .03, 1)) # Expected discounted utility after 1st cycle
```

```
[1] 0.9325
```

Discounted QALYs for each cycle

```
compute_qalys <- function(x, utility, dr = .03){  
  n_cycles <- nrow(x) - 1  
  pv(x %*% utility, dr, 0:n_cycles)  
}
```

```
dqalys_soc <- compute_qalys(x_soc, utility = utility)  
dqalys_new <- compute_qalys(x_new, utility = utility)
```

```
head(dqalys_soc)
```

```
[,1]  
0 1.0000000  
1 0.9325243  
2 0.8871161  
3 0.8484324  
4 0.8125404  
5 0.7784024
```

```
head(dqalys_new)
```

```
[,1]  
0 1.0000000  
1 0.9401942  
2 0.8980022  
3 0.8619435  
4 0.8285724  
5 0.7968343
```

Discounted cost for each cycle

```
compute_costs <- function(x, costs_medical, costs_treat, dr = .03){  
  
  n_cycles <- nrow(x) - 1  
  costs <- cbind(pv(x %*% costs_medical, dr, 0:n_cycles),  
                 pv(x %*% costs_treat, dr, 0:n_cycles))  
}  
  colnames(costs) <- c("medical", "treatment")  
  return(costs)  
}  
  
dcosts_soc <- compute_costs(x_soc, costs_medical, costs_treat_soc)  
dcosts_new <- compute_costs(x_new, costs_medical, costs_treat_new)
```

head(dcosts_soc)

	medical	treatment
0	2000.000	2000.000
1	2229.126	1937.864
2	2419.321	1876.527
3	2581.884	1816.350
4	2721.140	1757.443
5	2839.541	1699.850

head(dcosts_new)

	medical	treatment
0	2000.000	12000.00
1	2171.650	11631.84
2	2293.695	11270.64
3	2391.402	10917.83
4	2473.004	10573.78
5	2541.624	10238.54

Cost-effectiveness

```
(sum(dcosts_new[-1, ]) - sum(dcosts_soc[-1, ])) /  
(sum(dqalys_new[-1, ]) - sum(dqalys_soc[-1, ]))
```

[1] 122946.8

```
format_costs <- function(x) formatc(x, format = "d", big.mark = ",")  
format_qalys <- function(x) formatc(x, format = "f", digits = 2)
```

```
make_icer_tb1 <- function(costs0, costs1, qalys0, qalys1){
```

```
# Computations  
total_costs0 <- sum(costs0)  
total_costs1 <- sum(costs1)  
total_qalys0 <- sum(qalys0)  
total_qalys1 <- sum(qalys1)  
incr_total_costs <- total_costs1 - total_costs0  
inc_total_qalys <- total_qalys1 - total_qalys0  
icer <- incr_total_costs/inc_total_qalys
```

```
# Make table  
tibble(  
  `Costs` = c(total_costs0, total_costs1) %>%  
  `Strategy` = c("SOC", "New"),  
  format_costs(),  
  `QALYS` = c(total_qalys0, total_qalys1) %>%  
  format_qalys(),  
  `Incremental costs` = c("--", incr_total_costs %>% format_costs()),  
  `Incremental QALYS` = c("--", inc_total_qalys %>% format_qalys()),  
  `ICER` = c("--", icer) %>% format_costs()  
) %>%  
kable() %>%  
kable_styling() %>%  
footnote(general = "Costs and QALYS are discounted at 3% per annum.", footnote_as_chunk = TRUE)  
}
```

Cost-effectiveness

```
make_icer_tb1(costs0 = dcosts_soc[-1, ], costs1 = dcosts_new[-1, ],
               qalys0 = dqalys_soc[-1, ], qalys1 = dqalys_new[-1, ])
```

Strategy	Costs	QALYs	Incremental costs	Incremental QALYs	ICER
SOC	204,123	21.08	--	--	--
New	464,390	23.19	260,266	2.12	122,946

Note: Costs and QALYs are discounted at 3% per annum.

Steps

- Define transition matrix SOC
- Define transition matrix New treatment `apply_rr(p_soc, rr)`
- Define utility and cost values by health state
- Calculate health state probabilities over time `sim_markov_chain(x0, p, n_cycles)`
- Calculate expected QALYs and costs
 - `pv(z, dr, t)`
 - `compute_qalys(x, utility, dr)`
 - `compute_costs(x, costs_medical, costs_treat, dr)`
- Cost-effectiveness analysis `make_icer_tbl(costs0, costs1, qalys0, qalys1)`

Complete R script

```
01-markov-cohort.R x
Source on Save | Run | Source | ...
1 ## ---- Overview -----
2 ## @knitr R-packages
3 library("rcea")
4 library("knitr")
5 library("kableExtra")
6 library("magrittr")
7 library("tibble")
8
9 ## ---- Model parameters -----
10 ## @knitr transition-probabilities
11 p_hd <- .0002 # constant probability of dying when Healthy (all-cause mortality)
12 p_hs1 <- .15 # probability of becoming Sick when Healthy
13 p_s1h <- .05 # probability of becoming Healthy when Sick
14 p_s1s2 <- .0105 # probability of becoming Sicker when Sick
15 p_s1d <- .006 # constant probability of dying when Sick
16 p_s2d <- .02 # constant probability of dying when Sicker
17
18 ## @knitr transition-probability-complements
19 p_hh <- 1 - p_hs1 - p_hd
20 p_s1s1 <- 1 - p_s1h - p_s1s2 - p_s1d
21 p_s2s2 <- 1 - p_s2d
22
23 ## @knitr tpmatrix
24 p_soc <- matrix(
25   c(p_hh, p_hs1, 0, p_hd,
26     p_s1h, p_s1s1, p_s1s2, p_s1d,
27     0, 0, p_s2s2, p_s2d,
28     0, 0, 0, 1),
29   byrow = TRUE,
30   nrow = 4, ncol = 4
31 )
32 state_names <- c("H", "S1", "S2", "D")
33 colnames(p_soc) <- rownames(p_soc) <- state_names
34 print(p_soc)
35
36 ## @knitr apply_rr
37 apply_rr <- function(p, rr = .8){
38   p["H", "S1"] <- p["H", "S1"] * rr
39   p["H", "S2"] <- p["H", "S2"] * rr
40   p["H", "D"] <- p["H", "D"] * rr
41   p["H", "H"] <- 1 - sum(p["H", -1])
42
43   p["S1", "S2"] <- p["S1", "S2"] * rr
44   p["S1", "D"] <- p["S1", "D"] * rr
45   p["S1", "S1"] <- 1 - sum(p["S1", -2])
```

Tutorial

rcea **0.1.2**

Reference

Tutorials ▾

Slides



Simple Markov Cohort Model

2021-07-26

Source: vignettes/01-markov-cohort.Rmd

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Overview

Theory

Model parameters

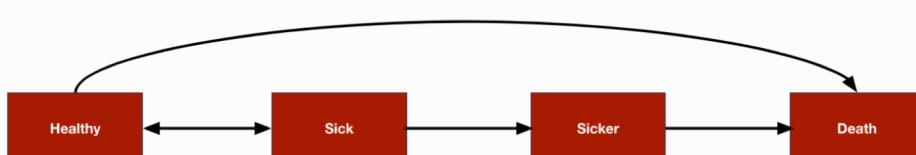
Simulation

<https://hesim-dev.github.io/rcea/articles/01-markov-cohort.html>

REFERRED TO AS A MARKOV COHORT MODEL. IN THIS TUTORIAL WE DEMONSTRATE IMPLEMENTATION WITH R OF THE SIMPLEST OF COST-EFFECTIVENESS MODELS, A LITTLE HOMOGENEOUS MODEL WITH TRANSITION PROBABILITIES THAT ARE CONSTANT OVER TIME. THE ENTIRE ANALYSIS CAN BE RUN USING **BASE R** (I.E., WITHOUT INSTALLING ANY PACKAGES). HOWEVER, WE WILL USE THE FOLLOWING PACKAGES TO CREATE A NICE LOOKING COST-EFFECTIVENESS TABLE.

```
library("rcea")
library("knitr")
library("kableExtra")
library("magrittr")
library("tibble")
```

As an example, we will consider the 4-state sick-sicker model that has been described in more detail by [Alarid-Escudero et al.](#) The model will be used to compare two treatment strategies, a “New” treatment and the existing “standard of care (SOC)”. The model consists of 4 health states. Ordered from worst to best to worst, they are: Healthy (*H*), Sick (*S₁*), Sicker (*S₂*), and Death (*D*). Possible transitions from each state are displayed in the figure below.



Exercise 1: Simple Markov Cohort model

- *Modify R-script “01-markov-cohort-R”*

- *Change relative risk*
 - *Change drug costs*
 - *Change utilities*
 - *Change follow-up time (i.e. number of cycles)*

`sim_markov_chain(x0, p, n_cycles)`

- *Run modified script*

Simple Markov cohort model – Incorporating probabilistic sensitivity analysis

Probabilistic sensitivity analysis

- The standard methodology for quantifying the impact of parameter uncertainty is probabilistic sensitivity analysis (PSA)
- Propagating uncertainty in the input parameters throughout the model by randomly sampling sets of input values from suitable probability distributions
- Probability distributions are determined according to the distributional properties of the statistical estimates, which, in turn, depend on the statistical techniques used and the distributions of the underlying data
- R is a natural programming language for performing PSA
- Random samples can be drawn from almost any probability distribution

Transition probabilities for SOC

- We assume that summary level data is available on transitions from the Healthy state ($n = 1000$), Sick state ($n = 1000$), and Sicker state ($n = 800$).

```
transitions_soc <- matrix(  
  c(848, 150, 0, 2,  
    500, 389, 105, 6,  
    0, 0, 784, 16,  
    0, 0, 0, 23),  
  nrow = 4, byrow = TRUE)  
  
state_names <- c("H", "S1", "S2", "D")  
colnames(transitions_soc) <- rownames(transitions_soc) <- tolower(state_names)
```

	h	s1	s2	d
h	848	150	0	2
s1	500	389	105	6
s2	0	0	784	16
d	0	0	0	23

- The transitions from each state to the other 4 states can be modeled using a Dirichlet distribution

Combining all model parameters

```
params <- list(
  alpha_soc = transitions_soc,
  lrr_mean = log(.8),
  lrr_lower = log(.71),
  lrr_upper = log(.9),
  c_medical = c(H = 2000, S1 = 4000, S2 = 15000, D = 0),
  c_soc = 2000,
  c_new = 12000,
  u_mean = c(H = 1, S1 = .75, S2 = 0.5, D = 0),
  u_se = c(H = 0, S1 = 0.03, S2 = 0.05, D = 0.0)
)
```

Simulation

- The simulation proceeds by
 1. randomly sampling the parameters from the probability distributions specified
 2. running the Markov model for each draw of the parameters
- The result is a draw from the probability distribution of each of the model outputs of interest (i.e., state probabilities, QALYs, and costs).

Sampling parameters

- While Base R can be used to draw samples of parameters, the functions `hesim::define_rng()` and `hesim::eval_rng()` simplify this process.
- Any random number generation function can be used inside the `define_rng()` block

```
rng_def <- define_rng({  
  
    lrr_se <- (lrr_upper - lrr_lower)/(2 * qnorm(.975))  
  
    list( # Parameters to return  
        p_soc = dirichlet_rng(alpha_soc),  
        rr_new = lognormal_rng(lrr_mean, lrr_se),  
        c_medical = gamma_rng(mean = c_medical, sd = c_medical),  
        c_soc = c_soc,  
        c_new = c_new,  
        u = beta_rng(mean = u_mean, sd = u_se)  
    )  
}, n = 1000)  
  
params_rng <- eval_rng(rng_def, params = params)
```

Sampling parameters

```
names(params_rng)
```

```
[1] "p_soc"      "rr_new"     "c_medical"  "c_soc"      "c_new"      "u"
```

```
head(as.matrix(params_rng$p_soc))
```

	h_h	h_s1	h_s2	h_d	s1_h	s1_s1	s1_s2	s1_d	s2_h	s2_s1
[1,]	0.8686001	0.1293872	0	0.002012649	0.5003043	0.3847703	0.10868135	0.006244105	0	0
[2,]	0.8474698	0.1499868	0	0.002543368	0.5146218	0.3964675	0.08375492	0.005155768	0	0
[3,]	0.8559237	0.1428611	0	0.001215126	0.5117574	0.3775261	0.10868021	0.002036291	0	0
[4,]	0.8550586	0.1429657	0	0.001975648	0.5139857	0.3777508	0.10303967	0.005223752	0	0
[5,]	0.8678962	0.1304462	0	0.001657694	0.5164343	0.3815376	0.09725638	0.004771722	0	0
[6,]	0.8530231	0.1459388	0	0.001038023	0.4991514	0.3750200	0.11846736	0.007361245	0	0
	s2_s2	s2_d	d_h	d_s1	d_s2	d_d				
[1,]	0.9786377	0.02136235	0	0	0	1				
[2,]	0.9871702	0.01282980	0	0	0	1				
[3,]	0.9786291	0.02137091	0	0	0	1				
[4,]	0.9809938	0.01900624	0	0	0	1				
[5,]	0.9793886	0.02061141	0	0	0	1				
[6,]	0.9750211	0.02497889	0	0	0	1				

Simulating the Markov model

- One way that a Markov simulation can be generalized is to store “input data” in an object, i.e. data frame.
- Input data might consist of
 - treatment strategies
 - patients and subgroups
 - For instance, if we were simulating different subgroups we might store the age and sex associated with the subgroup which could, in turn, be used as covariates in a statistical model.

```
data <- data.frame(  
  strategy = c("New", "SOC")  
)
```

	strategy
1	New
2	SOC

Simulating the Markov model – Create the function

- Set up a `sim_model()` function that runs the entire simulation.
 - Comprised of three smaller functions:
 - `sim_stateprobs()`
 - `compute_qalys()`
 - `compute_costs()`

Simulating the Markov model – Create the function

```
sim_stateprobs <- function(p0, rr, strategy, n_cycles){  
  
  rr <- ifelse(strategy == "New", rr, 1)  
  
  p <- tpmatrix(  
    C,           p0$h_s1 * rr,  p0$h_s2 * rr,  p0$h_d * rr,  
    p0$s1_h,     C,           p0$s1_s2 * rr,  p0$s1_d * rr,  
    p0$s2_h,     p0$s2_s1,   C,           p0$s2_d * rr,  
    0,           0,           0,           1  
  )  
  
  x <- sim_markov_chain(x0 = c(1, 0, 0, 0),  
                         p = matrix(as.matrix(p), ncol = 4, byrow = TRUE),  
                         n_cycles = n_cycles)  
  
  return(x)  
}
```

`hesim::tpmatrix()` makes it easy to define a transition probability matrix.

`C` denotes that a given element is the complement of all other elements in that row, ensuring that the probabilities sum to 1.

`sim_markov_chain()` is the function we created previously

Simulating the Markov model – Create the function

```
# QALYS
compute_qalys <- function(x, utility, dr = .03){

  n_cycles <- nrow(x) - 1
  pv(x %*% utility, dr, 0:n_cycles)
}
```

```
# Costs
compute_costs <- function(x, costs_medical, costs_treat, dr = .03){

  n_cycles <- nrow(x) - 1
  costs_treat <- c(rep(costs_treat, 3), 0)
  costs <- cbind(
    pv(x %*% costs_medical, dr, 0:n_cycles),
    pv(x %*% costs_treat, dr, 0:n_cycles)
  )
  colnames(costs) <- c("dcost_med", "dcost_treat")
  return(costs)
}
```

```

sim_model <- function(params_rng, data, n_cycles = 85, dr_qalys = .03, dr_costs = .03){
  PSA samples; tx strategy; no. cycles; discount rates
  # Initialize array of matrices
  n_samples <- attr(params_rng, "n")
  n_strategies <- nrow(data)
  out <- array(NA, dim = c(n_cycles + 1, 7, n_samples * n_strategies))
  dimnames(out) <- list(NULL, c("H", "S1", "S2", "D", "dqalys", "dcosts_med", "dcosts_treat"), NULL)

  # Run the simulation
  i <- 1
  for (s in 1:n_samples){ # Start PSA loop
    for (k in 1:n_strategies) { # Start treatment strategy loop
      x <- sim_stateprobs(p0 = params_rng$p_soc[s, ],
                           rr = params_rng$rr_new[s],
                           strategy = data$strategy[k],
                           n_cycles = n_cycles)
      dqalys <- compute_qalys(x, utility = unlist(params_rng$u[s]), dr = dr_qalys)
      dcosts <- compute_costs(x,
                               costs_medical = unlist(params_rng$c_medical[s]),
                               costs_treat = ifelse(data$strategy[k] == "SOC",
                                                    params_rng$c_soc,
                                                    params_rng$c_new), dr = dr_costs)
      out[, , i] <- cbind(x, dqalys, dcosts)
      i <- i + 1
    } # End treatment strategy loop
  } # End PSA loop

  # Store metadata and return
  attr(out, "n_samples") <- n_samples
  attr(out, "strategies") <- data$strategy
  return(out)
}

```

An array to store the output.
A series of matrices each with n_cycles rows and columns for each output.
There is one matrix for each parameter sample for the PSA and treatment strategy.

Simulates for each parameter sample and treatment strategy

- state probabilities (with sim_stateprobs())
- QALYs (with compute_qalys())
- costs (with compute_costs())

The number of parameter samples and the names of the treatment strategies are saved as attributes (i.e., metadata) to the array.

Simulating the Markov model

```
sim_out <- sim_model(params_rng, data = data)

head(sim_out[, , 1])
```

	H	S1	S2	D	dqalys	dcosts_med	dcosts_treat
[1,]	1.0000000	0.0000000	0.000000000	0.000000000	1.0000000	639.2051	12000.00
[2,]	0.8927749	0.1055827	0.000000000	0.001642364	0.9440327	968.8714	11631.35
[3,]	0.8498706	0.1371191	0.009363733	0.003646603	0.9027066	1072.4091	11269.90
[4,]	0.8273444	0.1453902	0.021361082	0.005904295	0.8667374	1105.0422	10916.86
[5,]	0.8113717	0.1463692	0.033882825	0.008376274	0.8332591	1114.3372	10572.54
[6,]	0.7976015	0.1450801	0.046273111	0.011045289	0.8013670	1114.9710	10236.97

Reorganize output

```
sim_out <- array_to_dt(sim_out)  
head(sim_out)
```

Convert a 3D array (faster to store data) to a data.table so we can summarize outcomes for each parameter sample and treatment strategy very quickly.

	cycle	strategy	sample	H	S1	S2	D	dqalys	dcosts_med	dcosts_treat
1:	0	New	1	1.0000000	0.0000000	0.000000000	0.000000000	1.0000000	639.2051	12000.00
2:	1	New	1	0.8927749	0.1055827	0.000000000	0.001642364	0.9440327	968.8714	11631.35
3:	2	New	1	0.8498706	0.1371191	0.009363733	0.003646603	0.9027066	1072.4091	11269.90
4:	3	New	1	0.8273444	0.1453902	0.021361082	0.005904295	0.8667374	1105.0422	10916.86
5:	4	New	1	0.8113717	0.1463692	0.033882825	0.008376274	0.8332591	1114.3372	10572.54
6:	5	New	1	0.7976015	0.1450801	0.046273111	0.011045289	0.8013670	1114.9710	10236.97

Cost-effectiveness output

```
ce_sim <- sim_out[cycle != 0,
                  .(dqalys = sum(dqalys),
                    dcots = sum(dcots_med) + sum(dcots_treat)),
                  by = c("sample", "strategy")]
ce_sim
```

```
      sample strategy    dqalys    dcots
1:      1       New 23.30567 372636.3
2:      1       SOC 21.36614 103030.7
3:      2       New 23.49382 380159.8
4:      2       SOC 22.71959 104017.1
5:      3       New 23.35243 463270.5
---
1996:   998       SOC 20.63295 166641.1
1997:   999       New 21.82111 356686.6
1998:   999       SOC 19.58578 104643.0
1999:  1000       New 23.03594 559476.0
2000:  1000       SOC 21.79923 320811.6
```

Save for later

```
saveRDS(ce_sim, file = "markov-cohort-ce_sim.rds")
```

Cost-effectiveness output

```
ce_sim_wider <- dcast(ce_sim, sample ~ strategy,  
                      value.var = c("dqalys", "dcosts"))
```

```
ce_sim_wider
```

	sample	dqalys_New	dqalys_SOC	dcosts_New	dcosts_SOC
1:	1	23.30567	21.36614	372636.3	103030.72
2:	2	23.49382	22.71959	380159.8	104017.07
3:	3	23.35243	21.81066	463270.5	207518.50
4:	4	23.49622	21.45324	680491.2	474308.21
5:	5	24.11580	22.73777	492869.3	239480.34

996:	996	23.05054	20.86090	578855.4	336586.31
997:	997	24.22340	22.48457	379255.5	93578.69
998:	998	22.22956	20.63295	452278.4	166641.12
999:	999	21.82111	19.58578	356686.6	104642.99
1000:	1000	23.03594	21.79923	559476.0	320811.62

Cost-effectiveness output

```
ce_sim_wider[, idcosts := dcosts_New - dcosts_SOC]  
ce_sim_wider[, idqalys := dqalys_New - dqalys_SOC]
```

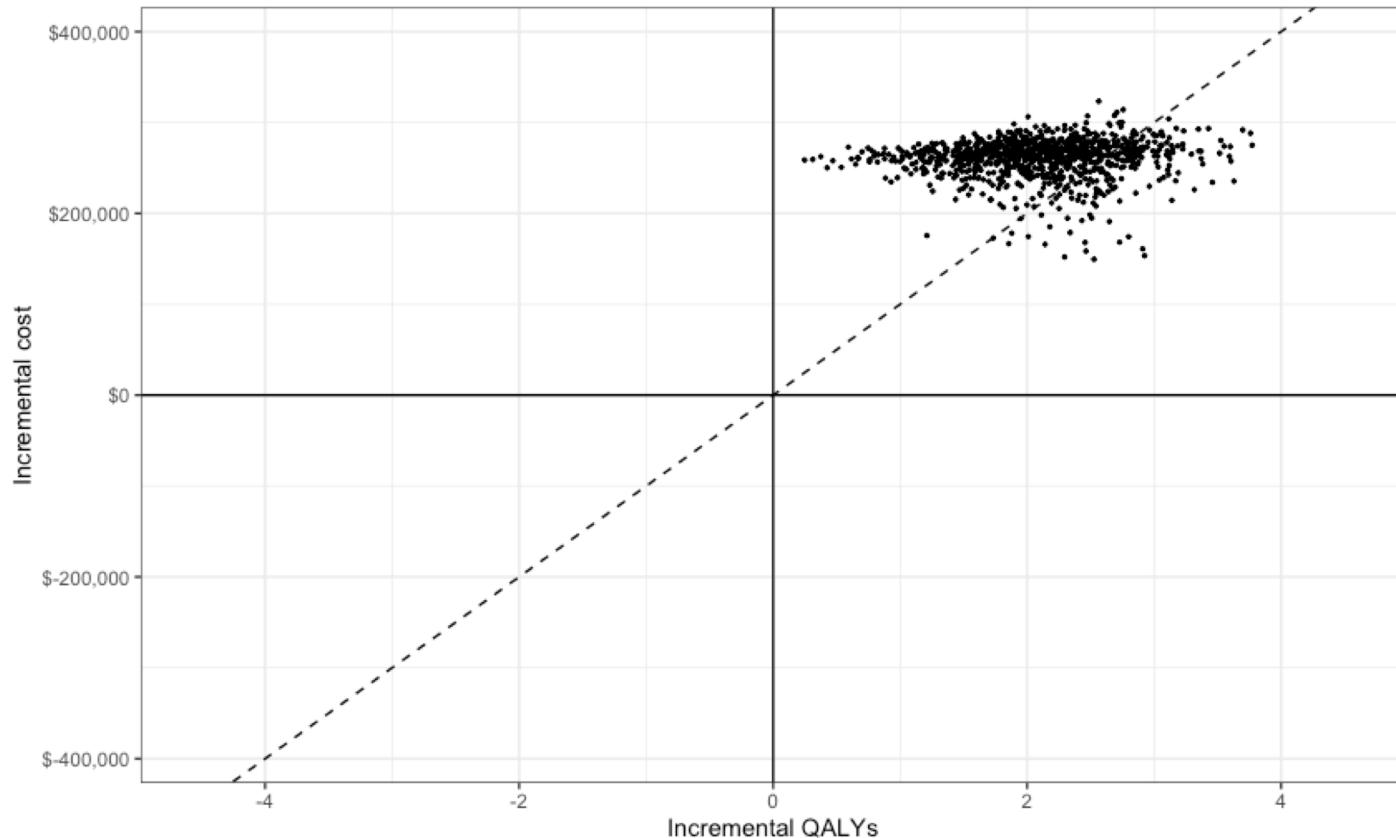
	sample	dqalys_New	dqalys_SOC	dcosts_New	dcosts_SOC	idcosts	idqalys
1:	1	23.30567	21.36614	372636.3	103030.72	269605.6	1.9395308
2:	2	23.49382	22.71959	380159.8	104017.07	276142.7	0.7742344
3:	3	23.35243	21.81066	463270.5	207518.50	255752.0	1.5417635
4:	4	23.49622	21.45324	680491.2	474308.21	206183.0	2.0429732
5:	5	24.11580	22.73777	492869.3	239480.34	253388.9	1.3780308

996:	996	23.05054	20.86090	578855.4	336586.31	242269.1	2.1896429
997:	997	24.22340	22.48457	379255.5	93578.69	285676.8	1.7388263
998:	998	22.22956	20.63295	452278.4	166641.12	285637.3	1.5966079
999:	999	21.82111	19.58578	356686.6	104642.99	252043.6	2.2353267
1000:	1000	23.03594	21.79923	559476.0	320811.62	238664.4	1.2367111

```
ce_sim_wider[, .(icer = mean(idcosts)/mean(idqalys))]
```

	icer
1:	125028.7

Cost-effectiveness plane



Steps

- Define uncertainty for all model input parameters
- Sampling parameter values `hesim::define_rng()`, `hesim::eval_rng()`
- Define treatment strategies and population (data)
- Simulating the Markov model

```
sim_model(params_rng, data, n_cycles, dr_qalys, dr_costs)
sim_stateprobs(po, rr, strategy, n_cycles)
compute_qalys(x, utility, dr)
compute_costs(x, cost_medical, costs_treat, dr)
```

- Reorganize output `rbind_array()`, `array_to_dt()`
- Cost-effectiveness analysis

Complete R script

```
02-markov-cohort-psa.R   
Source on Save | Run | Source |   
1 ## ---- Overview -----  
2 ## @knitr R-packages  
3 library("rcaea")  
4 library("hesim")  
5 library("data.table")  
6 library("magrittr")  
7 library("ggplot2")  
8  
9 ## ---- Model parameters -----  
10 ## @knitr tpmatrix  
11 transitions_soc <- matrix(  
12   c(848, 150, 0, 2,  
13     500, 389, 105, 6,  
14     0, 0, 784, 16,  
15     0, 0, 0, 23),  
16   nrow = 4, byrow = TRUE)  
17 state_names <- c("H", "S1", "S2", "D")  
18 colnames(transitions_soc) <- rownames(transitions_soc) <- tolower(state_names)  
19  
20 ## @knitr all-parameters  
21 params <- list(  
22   alpha_soc = transitions_soc,  
23   lrr_mean = log(.8),  
24   lrr_lower = log(.71),  
25   lrr_upper = log(.9),  
26   c_medical = c(H = 2000, S1 = 4000, S2 = 15000, D = 0),  
27   c_soc = 2000,  
28   c_new = 12000,  
29   u_mean = c(H = 1, S1 = .75, S2 = 0.5, D = 0),  
30   u_se = c(H = 0, S1 = 0.03, S2 = 0.05, D = 0.0)  
31 )  
32  
33 ## ---- Simulation -----  
34 ## @knitr sample-parameters  
35 rng_def <- define_rng({  
36   lrr_se <- (lrr_upper - lrr_lower)/(2 * qnorm(.975)) # Local object  
37   # not returned  
38   list( # Parameters to return  
39     p_soc = dirichlet_rng(alpha_soc),  
40     rr_new = lognormal_rng(lrr_mean, lrr_se),  
41     c_medical = gamma_rng(mean = c_medical, sd = c_medical),  
42     c_soc = c_soc,  
43     c_new = c_new,  
44     u = beta_rng(mean = u_mean, sd = u_se)  
45   )  
46 sim_model(params_rng, data, n_cycles, dr_qalys, dr_costs) #
```

Tutorial

rcea **0.1.2**

Reference

Tutorials ▾

Slides



2021-07-26

Source: vignettes/02-markov-cohort-psa.Rmd

Overview

Probabilistic sensitivity analysis (PSA) is used to quantify the impact of parameter uncertainty on the uncertainty of model outputs. PSA is typically performed via a simulation approach whereby the model parameters are randomly sampled from suitable probability distributions

<https://hesim-dev.github.io/rcea/articles/02-markov-cohort-psa.html>

```
library("rcea")
library("hesim")
library("data.table")
library("magrittr")
library("ggplot2")
```



Model parameters

Transition probabilities for SOC

The probability distribution used for transition probabilities will depend on the underlying data. In this case, we assume that summary level data is available on transitions from the Healthy state ($n = 900$), Sick state ($n = 900$), and Sicker state ($n = 800$). The transitions from each state to the other 4 states can be modeled using a Dirichlet distribution (see Appendix).

```
transitions_soc <- matrix(
  c(848, 150, 0, 2,
```

Exercise 2: Incorporating probabilistic sensitivity analysis

- *Modify R-script “02-markov-cohort-psa.R”*
 - *Reduce sample size of data for transition matrix by 50%*
 - *Increase confidence interval for relative risk*
- *Run modified script*

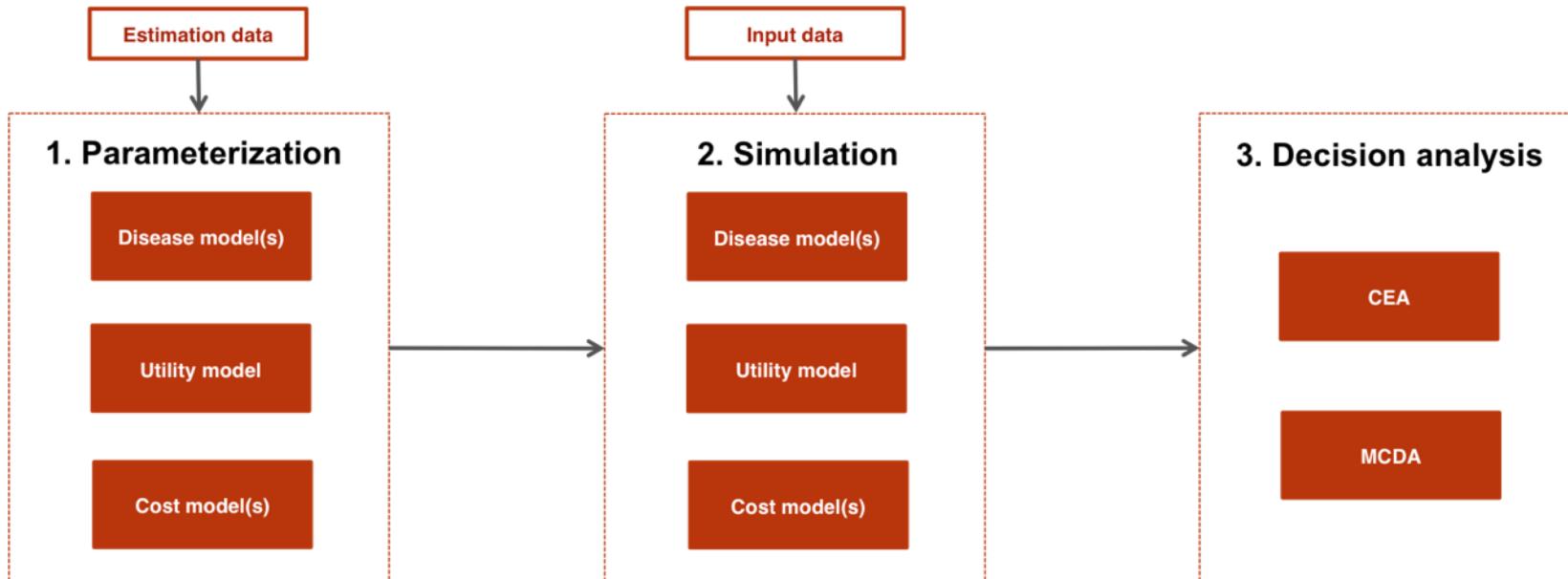
Simple Markov cohort model with hesim



What is hesim?

- A modular and computationally efficient R package for building simulation models for economic evaluation
- Supports both cohort and individual-level models, encompassing Markov (time-homogeneous and time-inhomogeneous) and semi-Markov processes
 - cohort discrete time state transition models (cDTSTM)
 - individual-level continuous time state transition models (iCTSTM)
 - n-state partitioned survival models (PSM)
- Parameterization by fitting a statistical model in R or by using estimates from external sources
- Nearly all simulation code written in C++ under the hood, but you don't need to know C++ to use it!

hesim modeling process



Economic models with hesim

1. Model set-up
2. Parameters
3. Simulation
 - a. Construction of model
 - b. Simulation of outcomes
4. Cost-effectiveness analysis

Model setup

- Define target population and intervention strategies

```
strategies <- data.frame(  
  strategy_id = 1:2,  
  strategy_name = c("SOC", "New")  
)  
patients <- data.frame(  
  patient_id = 1,  
  age = 25  
)  
hesim_dat <- hesim_data(  
  strategies = strategies,  
  patients = patients  
)  
  
print(hesim_dat)
```

```
$strategies  
  strategy_id strategy_name  
1           1          SOC  
2           2          New  
  
$patients  
  patient_id age  
1           1    25  
  
attr(,"class")  
[1] "hesim_data"
```

```
Tabs <- get_labels(hesim_dat)
```

Model parameters

- Same list of parameters as used before

```
params <- list(  
  alpha_soc = transitions_soc,  
  lrr_mean = log(.8),  
  lrr_lower = log(.71),  
  lrr_upper = log(.9),  
  c_medical = c(H = 2000, S1 = 4000, S2 = 15000),  
  c_soc = 2000,  
  c_new = 12000,  
  u_mean = c(H = 1, S1 = .75, S2 = 0.5),  
  u_se = c(H = 0, S1 = 0.03, S2 = 0.05)  
)
```

Model parameters – Random number generation (for PSA)

```
rng_def <- define_rng({  
  lrr_se <- (lrr_upper - lrr_lower)/(2 * qnorm(.975))  
  
  list( # Parameters to return  
    p_soc = dirichlet_rng(alpha_soc),  
    rr_new = lognormal_rng(lrr_mean, lrr_se),  
    c_medical = gamma_rng(mean = c_medical, sd = c_medical),  
    c_soc = c_soc,  
    c_new = c_new,  
    u = beta_rng(mean = u_mean, sd = u_se)  
  )  
}, n = 1000)
```

Model parameters – Transformed parameters

- Typically underlying parameters (`params`) are transformed (`tparams`) into more relevant parameters for the simulation
 - e.g., predicting an element of a transition probability matrix as a function of the treatment strategy
- We previously did this in the base R PSA example...

```

sim_model <- function(params_rng, data, n_cycles = 85, dr_qalys = .03, dr_costs = .03){

  # Initialize array of matrices
  n_samples <- attr(params_rng, "n")
  n_strategies <- nrow(data)
  out <- array(NA, dim = c(n_cycles + 1, 7, n_samples * n_strategies))
  dimnames(out) <- list(NULL, c("H", "S1", "S2", "D", "dqalys", "dcosts_med", "dcosts_treat"), NULL)

  # Run the simulation
  i <- 1
  for (s in 1:n_samples){ # Start PSA loop
    for (k in 1:n_strategies) { # Start treatment strategy loop
      x <- sim_stateprobs(p0 = params_rng$p_soc[s, ],
                           rr = params_rng$rr_new[s],
                           strategy = data$strategy[k],
                           n_cycles = n_cycles)
      dqalys <- compute_qalys(x, utility = unlist(params_rng$u[s]), dr = dr_qalys)
      dcosts <- compute_costs(x,
                               costs_medical = unlist(params_rng$c_medical[s]),
                               costs_treat = ifelse(data$strategy[k] == "SOC",
                                                    params_rng$c_soc,
                                                    params_rng$c_new), dr = dr_costs)
      out[, , i] <- cbind(x, dqalys, dcosts)
      i <- i + 1
    } # End treatment strategy loop
  } # End PSA loop

  # Store metadata and return
  attr(out, "n_samples") <- n_samples
  attr(out, "strategies") <- data$strategy
  return(out)
}

sim_out <- sim_model(params_rng, data = data)
sim_out <- array_to_dt(sim_out)

```

```
sim_stateprobs <- function(p0, rr, strategy, n_cycles){

  rr <- ifelse(strategy == "New", rr, 1)

  p <- tpmatrix(
    C,           p0$h_s1 * rr,  p0$h_s2 * rr,  p0$h_d * rr,
    p0$s1_h,     C,           p0$s1_s2 * rr,  p0$s1_d * rr,
    p0$s2_h,     p0$s2_s1,   C,           p0$s2_d * rr,
    0,           0,           0,           1
  )

  x <- sim_markov_chain(x0 = c(1, 0, 0, 0),
                        p = matrix(as.matrix(p), ncol = 4, byrow = TRUE),
                        n_cycles = n_cycles)

  return(x)
}
```

Model parameters – Transformed parameters

- A `define_tparams()` block in `hesim` does the same thing, but most of the implementation is done for you (efficiently)
- A `define_tparams()` block returns:
 - `tpmatrix`: The transition probability matrix
 - `utility`: Utility assigned to each health state
 - `costs`: Costs assigned to each health state for each cost category
- All parameters are “transformed” using:
 1. Columns of input data
 2. Parameters returned by `define_rng()`

Model parameters – Transformed parameters

- *Input data* (treatment strategies and patients) can be generated using `expand()`¹

```
input_data <- expand(hesim_dat, by = c("strategies", "patients"))

head(input_data)
```

```
  strategy_id patient_id strategy_name age
1:          1          1           SOC  25
2:          2          1           New  25
```

¹Could also be expanded by time intervals in a time-inhomogeneous model

Model parameters – Transformed parameters

- You write mathematical expressions
- Vectorized over PSA iterations and input data rows

```
tparams_def <- define_tparams({  
  # The treatment effect (relative risk) varies by  
  # strategies (SOC is the reference strategy)  
  
  rr <- ifelse(strategy_name == "SOC", 1, rr_new)  
  
  list(  
    tpmatrix = tpmatrix(  
      C, p_soc$h_s1 * rr, p_soc$h_s2 * rr, p_soc$h_d * rr,  
      p_soc$s1_h, C, p_soc$s1_s2 * rr, p_soc$s1_d * rr,  
      p_soc$s2_h, p_soc$s2_s1, C, p_soc$s2_d * rr,  
      0, 0, 0, 1  
    ),  
    utility = u,  
    costs = list(  
      treatment = ifelse(strategy_name == "SOC", c_soc, c_new),  
      medical = c_medical  
    )  
  )  
})
```

Annotations:

- rr <- ifelse(strategy_name == "SOC", 1, rr_new)
 - rr: Input data
 - 1: Parameter
- tpmatrix(
 - C: Parameter
 - p_soc\$h_s1 * rr: Parameter (defined above)
 - p_soc\$h_s2 * rr: Parameter (defined above)
 - p_soc\$h_d * rr: Parameter (defined above)
 - p_soc\$s1_h: Parameter
 - C: Parameter (defined above)
 - p_soc\$s1_s2 * rr: Parameter (defined above)
 - p_soc\$s1_d * rr: Parameter (defined above)
 - p_soc\$s2_h: Parameter
 - p_soc\$s2_s1: Parameter
 - C: Parameter (defined above)
 - p_soc\$s2_d * rr: Parameter (defined above)
 - 0: Input data
 - 0: Input data
 - 0: Input data
 - 1: Parameter
- utility = u: Parameter
- costs = list(
 - treatment = ifelse(strategy_name == "SOC", c_soc, c_new): Input data
 - medical = c_medical: Input data
-)

tpmatrix() is a powerful function for creating/storing transition probabilities that vary across PSA samples, strategies, subgroups, and time intervals

C denotes the “complement”, ensuring that the probabilities in a row sum to 1

Simulation - Construct the model

- Combine the underlying parameters with the expressions for random number generation and parameter transformation

```
mod_def <- define_model(tparams_def = tparams_def,  
                        rng_def = rng_def,  
                        params = params)
```

- A economic model (of class CohortDtstm) can be created from a defined model (of class model_def) and data using the generic function `create_CohortDtstm()`

```
econmod <- create_CohortDtstm(mod_def, input_data)
```

- This object consists of a
 - transition model for simulating transition probabilities with `$sim_stateprobs()`
 - a utility model for simulating quality-adjusted life-years with `$sim_qalys()`
 - a set of cost models (for each cost category) for simulating costs with `$sim_costs()`

Simulation – Simulating outcomes

■ Health state probabilities

```
econmod$sim_stateprobs(n_cycles = 85)
```

	sample	strategy_id	patient_id	grp_id	state_id	t	prob
1:	1		1	1		1 0	1.0000000
2:	1		1	1		1 1	0.8466964
3:	1		1	1		1 2	0.7929241
4:	1		1	1		1 3	0.7652357
5:	1		1	1		1 4	0.7446251
6:	1		1	1		1 5	0.7261680

■ QALYs

```
econmod$sim_qalys(  
  dr = 0.03, lys = TRUE,  
  integrate_method = "riemann_right"  
)
```

	sample	strategy_id	patient_id	grp_id	state_id	dr	qalys	lys
1:	1		1	1		1 0.03	14.604941	14.604941
2:	1		1	1		2 0.03	2.718281	3.660628
3:	1		1	1		3 0.03	2.595318	7.331531
4:	1	2		1	1	1 0.03	17.633196	17.633196
5:	1	2		1	1	2 0.03	2.632315	3.544860
6:	1	2		1	1	3 0.03	2.083739	5.886368

■ Costs

```
econmod$sim_costs(  
  dr = 0.03,  
  integrate_method = "riemann_right"  
)
```

	sample	strategy_id	patient_id	grp_id	state_id	dr	category	costs
1:	1		1	1		1 0.03	treatment	29209.882
2:	1		1	1		2 0.03	treatment	7321.257
3:	1		1	1		3 0.03	treatment	14663.061
4:	1	2		1	1	1 0.03	treatment	211598.347
5:	1	2		1	1	2 0.03	treatment	42538.323
6:	1	2		1	1	3 0.03	treatment	70636.410

Cost-effectiveness analysis

- CEAs can be performed directly from the simulation output with **hesim**.
- First we need to aggregate (i.e., "summarize") costs and QALYs across health states

```
ce_sim <- econmod$summarize()
```

\$costs						
	category	dr	sample	strategy_id	costs	grp_id
1:	treatment	0.03	1		1 51194.20	1
2:	treatment	0.03	1		2 324773.08	1
3:	treatment	0.03	2		1 48720.41	1
4:	treatment	0.03	2		2 316828.55	1
5:	treatment	0.03	3		1 51943.80	1

5996:	total	0.03	998		2 402500.90	1
5997:	total	0.03	999		1 197991.68	1
5998:	total	0.03	999		2 461755.98	1
5999:	total	0.03	1000		1 203541.38	1
6000:	total	0.03	1000		2 461558.77	1

\$qalys						
	dr	sample	strategy_id	qalys	grp_id	
1:	0.03	1		1 19.91854	1	
2:	0.03	1		2 22.34925	1	
3:	0.03	2		1 19.87522	1	
4:	0.03	2		2 22.71959	1	
5:	0.03	3		1 21.39403	1	

1996:	0.03	998		2 23.08819	1	
1997:	0.03	999		1 21.25388	1	
1998:	0.03	999		2 22.98019	1	
1999:	0.03	1000		1 20.75811	1	
2000:	0.03	1000		2 23.84501	1	


```
attr(,"class")
[1] "ce"
```

Save for later

```
saveRDS(ce_sim, file = "markov-cohort-hesim-ce_sim.rds")
saveRDS(hesim_dat, file = "markov-cohort-hesim_data.rds")
```

Cost-effectiveness analysis

- Here, we will consider a pairwise comparison between the new treatment and SOC with the `cea_pw()` function

```
cea_pw_out <- cea_pw(ce_sim, comparator = 1,  
                      dr_qalys = 0.03, dr_costs = 0.03,  
                      k = seq(0, 25000, 500))
```

- Although `cea_pw()` allows users to summarize output from a PSA we will just create an ICER table using means for now

```
format(icer(cea_pw_out, k = 50000, labels = 1abs))
```

	Outcome	New
1: Incremental QALYs		2.07 (1.05, 3.13)
2: Incremental costs	257,297	(203,094, 284,985)
3: Incremental NMB	-153,833	(-210,941, -77,824)
4: ICER		124,342

Steps with `hesim`

1. Model set-up

- Specify the treatment strategies and target population(s)
 - `hesim_data()`

2. Parameters

- Estimate or define the parameters of the economic model
 - `define_rng()`, `define_tparams()`

3. Simulation

a. Construction of model

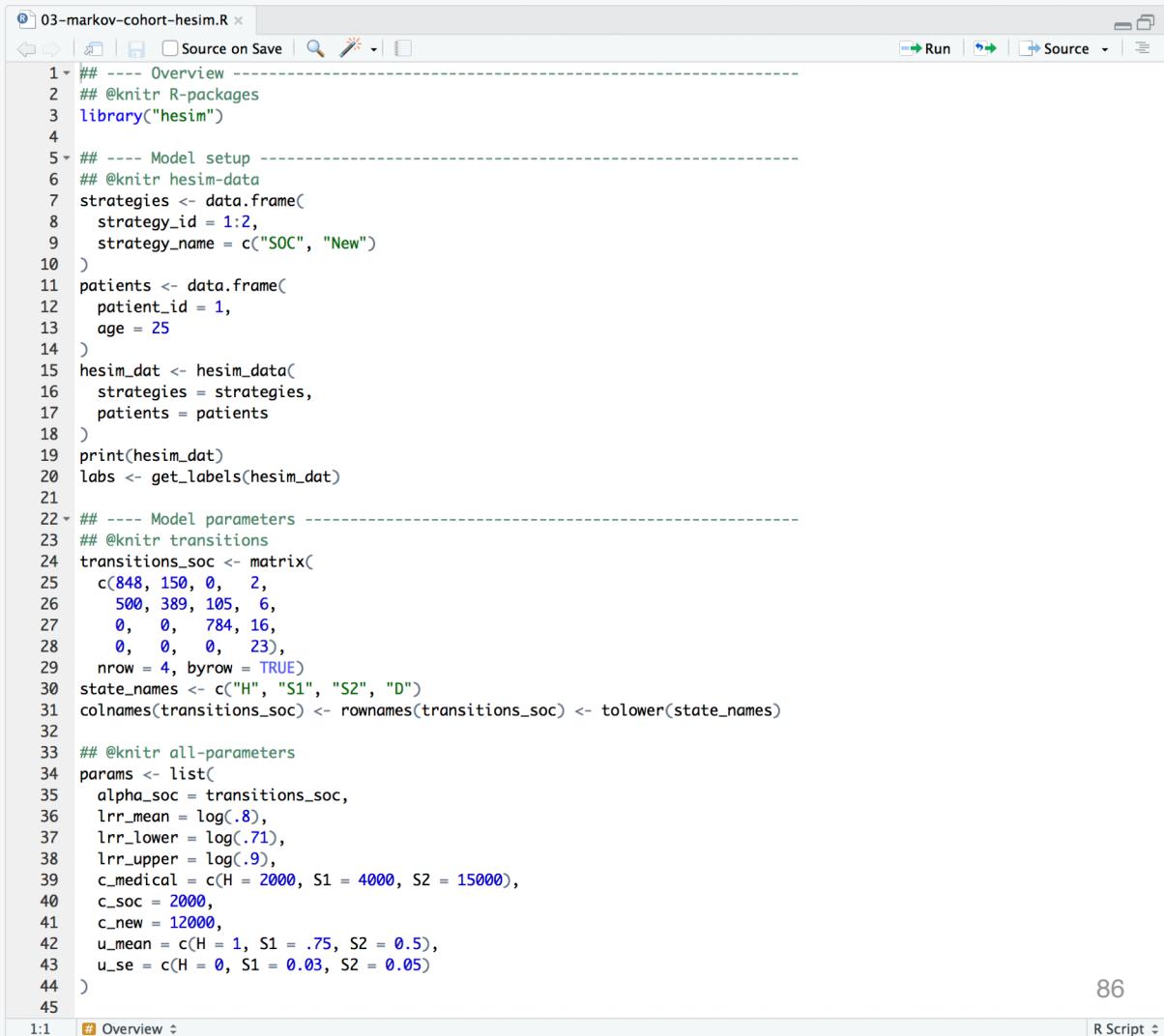
- Create an economic model—consisting of separate statistical models for disease progression, costs, and utilities—that simulate outcomes as a function of *input data* (derived from Step 1) and *parameters* (from Step 2)
- `define_model()`, `create_CohortDtsim()`

b. Simulation of outcomes

- Simulate outcomes (disease progression, costs, and quality-adjusted life-years (QALYs)) using the model constructed in Step 3
- `$sim_stateprobs()`, `$sim_qalys()`, `$sim_costs()`

4. Cost-effectiveness analysis

Complete R script



```
R 03-markov-cohort-hesim.R x
Source on Save | Run | Source | Overview | Help
1 ## ---- Overview -----
2 ## @knitr R-packages
3 library("hesim")
4
5 ## ---- Model setup -----
6 ## @knitr hesim-data
7 strategies <- data.frame(
8   strategy_id = 1:2,
9   strategy_name = c("SOC", "New")
10 )
11 patients <- data.frame(
12   patient_id = 1,
13   age = 25
14 )
15 hesim_dat <- hesim_data(
16   strategies = strategies,
17   patients = patients
18 )
19 print(hesim_dat)
20 labs <- get_labels(hesim_dat)
21
22 ## ---- Model parameters -----
23 ## @knitr transitions
24 transitions_soc <- matrix(
25   c(848, 150, 0, 2,
26     500, 389, 105, 6,
27     0, 0, 784, 16,
28     0, 0, 0, 23),
29   nrow = 4, byrow = TRUE)
30 state_names <- c("H", "S1", "S2", "D")
31 colnames(transitions_soc) <- rownames(transitions_soc) <- tolower(state_names)
32
33 ## @knitr all-parameters
34 params <- list(
35   alpha_soc = transitions_soc,
36   lrr_mean = log(.8),
37   lrr_lower = log(.71),
38   lrr_upper = log(.9),
39   c_medical = c(H = 2000, S1 = 4000, S2 = 15000),
40   c_soc = 2000,
41   c_new = 12000,
42   u_mean = c(H = 1, S1 = .75, S2 = 0.5),
43   u_se = c(H = 0, S1 = 0.03, S2 = 0.05)
44 )
45
# Overview
```

Tutorial

rcea **0.1.2**

Reference

Tutorials ▾

Slides



Markov Cohort Model with hesim

2021-07-26

<https://hesim-dev.github.io/rcea/articles/03-markov-cohort-hesim.html>

Overview

This tutorial repeats the probabilistic sensitivity analysis (PSA) of the Markov cohort model simulation performed in the [previous tutorial](#) using `hesim`. We utilize the cohort discrete time state transition model (`cDTSTM`) class, which is another name for a (time-homogeneous or time-inhomogeneous) Markov cohort model.

More information about `hesim` can be found by visiting the [package website](#). We recommend reading the "Articles"—starting with the "[Introduction to hesim](#)"—to learn more. Economic models can, in general, be simulated with the following steps:

1. **Model setup:** Specify the treatment strategies, target population(s), and model structure.
2. **Parameters:** Estimate or define the parameters of the economic model.
3. **Simulation:**
 - a. **Construction of model:** Create an economic model—consisting of separate statistical models for disease progression, costs, and utilities—that simulate outcomes as a function of *input data* (derived from Step 1) and *parameters* (from Step 2).
 - b. **Simulation of outcomes:** Simulate outcomes (disease progression, costs, and quality-adjusted life-years (QALYs)) using the model constructed in Step 3.

This analysis can be performed using the `hesim` package alone.

```
library("hesim")
```

Model setup

Before beginning an analysis, it is necessary to define the treatment strategies of interest and the the target population of interest. We

Contents

Overview

Simulation

Cost-effectiveness analysis

Exercise 3: Markov cohort model with hesim

- *Modify R-script “03-markov-cohort-hesim.R”*
 - Increase confidence interval for relative risk
 - Modify the mean health state utility value
 - Remove impact of the intervention on transitions from “healthy” to “sick”, “sicker”, and “death” (row 72)

- *Run modified script*

```
tparams_def <- define_tparams({  
  ## The treatment effect (relative risk) is transformed so that it varies by  
  ## strategies (SOC is the reference strategy)  
  rr <- ifelse(strategy_name == "SOC", 1, rr_new)  
  
  list(  
    tpmatrix = tpmatrix(  
      C, p_soc$h_s1 * rr, p_soc$h_s2 * rr, p_soc$h_d * rr,  
      p_soc$s1_h, C, p_soc$s1_s2 * rr, p_soc$s1_d * rr,  
      p_soc$s2_h, p_soc$s2_s1, C, p_soc$s2_d * rr,  
      0, 0, 0, 1  
    ),  
    utility = u,  
    costs = list(  
      treatment = ifelse(strategy_name == "SOC", c_soc, c_new),  
      medical = c_medical  
    )  
  )  
})
```

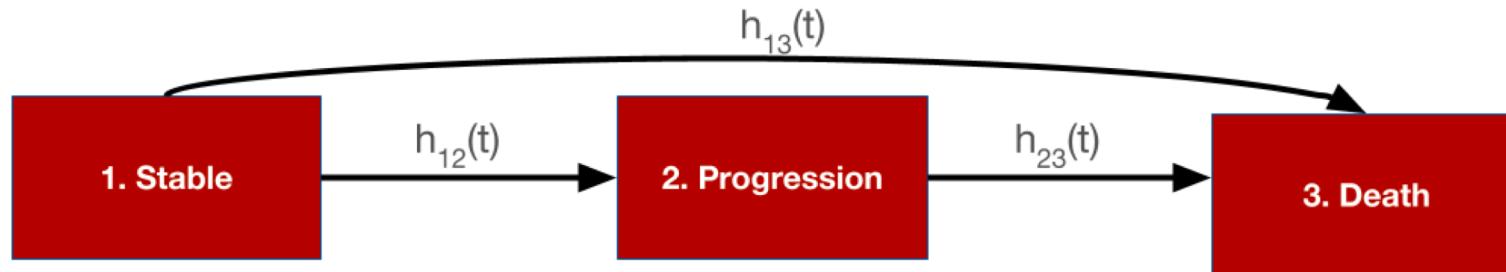
Semi-Markov multi-state model

Semi-Markov multi-state model

- Transition rates can depend on time in intermediate health states (unlike in a Markov model)
- Can only be simulated in a general manner using individual patient simulation (IPS)
- IPS is performed most efficiently using a **continuous time state transition model (CTSTM)**
- Ideally parameterizing by fitting a multi-state model

Semi-Markov multi-state model

Clock reset



Economic models with **hesim**

1. Model set-up
2. Parameters
3. Simulation
 - a. Construction of model
 - b. Simulation of outcomes
4. Cost-effectiveness analysis

Model setup

- The transitions of a multi-state model in **hesim** must be characterized by a matrix where each element denotes a transition from a row to a column

```
tmat <- rbind(  
  c(NA, 1, 2),  
  c(NA, NA, 3),  
  c(NA, NA, NA)  
)  
colnames(tmat) <- rownames(tmat) <- c("Stable", "Progression", "Dead")  
  
print(tmat)
```

	Stable	Progression	Dead
Stable	NA	1	2
Progression	NA	NA	3
Dead	NA	NA	NA

Model setup

```
n_patients <- 1000

patients <- data.table(
  patient_id = 1:n_patients,
  age = rnorm(n_patients, mean = 45, sd = 7),
  female = rbinom(n_patients, size = 1, prob = .51)
)

strategies <- data.frame(
  strategy_id = 1:2,
  strategy_name = c("SOC", "New")
)
n_strategies <- nrow(strategies)

states <- data.table(
  state_id = c(1, 2),
  state_name = c("Stable", "Progression")
)
n_states <- nrow(states)

hesim_dat <- hesim_data(
  strategies = strategies,
  patients = patients,
  states = states
)
```

As in the cohort model, we must specify the target population and treatment strategies of interest

In an IPS we simulate many patients and then average outcomes across the simulated patients. 1,000 simulated patients should produce reasonably stable results

We also explicitly define the (non-death) health states, which we will use to model utility and costs

We always combine this information into one object

Model setup

```
print(hesim_dat)
```

```
$strategies
  strategy_id strategy_name
1                 1             SOC
2                 2            New

$patients
  patient_id    age female
1:          1 42.71774     0
2:          2 48.86723     0
3:          3 40.27539     1
4:          4 46.50052     1
5:          5 47.17538     1
---
996:        996 43.22279     0
997:        997 54.22166     0
998:        998 37.27172     0
999:        999 45.20616     0
1000:       1000 39.15981    1

$states
  state_id state_name
1:        1      Stable
2:        2 Progression

attr(,"class")
[1] "hesim_data"
```

```
Tabs <- get_labels(hesim_dat)
```

Parameter estimation

- In the cohort examples, we used parameter estimates from the literature
 - We will continue to do this for utility and costs
- However, in an ideal scenario, we would estimate parameters ourselves using patient-level data
 - We will fit a multi-state model in this manner by estimating transition specific hazards using the R package **flexsurv**

Parameter estimation – Multi-state model

- Multi-state models can be fit by:
 - Estimating a joint survival model with interaction terms for different transition
 - Fitting separate survival models for each transition
(Method used here)

Parameter estimation – Multi-state model

■ Dataset

```
      from      to strategy_name female      age patient_id time_start
1: Stable Progression      New 0 59.85813 1 0.000000
2: Stable Death            New 0 59.85813 1 0.000000
3: Progression Death      New 0 59.85813 1 2.420226
4: Stable Progression      New 0 62.57282 2 0.000000
5: Stable Death            New 0 62.57282 2 0.000000

      time_stop status transition_id strategy_id      time
1: 2.420226    1           1          3 2.420226
2: 2.420226    0           2          3 2.420226
3: 14.620258   1           3          3 12.200032
4: 7.497464    0           1          3 7.497464
5: 7.497464    0           2          3 7.497464
```

■ Estimate parameters

```
wei_fits <- vector(length = 3, mode = "list")

for (i in 1:3){ # 3 possible transitions
  wei_fits[[i]] <- flexsurvreg(
    Surv(time, status) ~ strategy_name + female,
    data = data,
    subset = (transition_id == i) ,
    dist = "weibull")
}

wei_fits <- flexsurvreg_list(wei_fits)
```

Parameter estimation – Multi-state model

Stable -> Progression

```
[[1]]
Call:
flexsurvreg(formula = Surv(time, status) ~ strategy_name + female,
  data = data, subset = (transition_id == i), dist = "weibull")

Estimates:
          data mean   est     L95%    U95%    se   exp(est)   L95%    U95%
shape        NA  2.0152  1.9226  2.1124  0.0484      NA       NA       NA
scale        NA  7.1668  6.7796  7.5762  0.2031      NA       NA       NA
strategy_nameNew 0.5281  0.2745  0.2131  0.3360  0.0314  1.3159  1.2375  1.3994
female       0.4947 -0.1902 -0.2518 -0.1286  0.0314  0.8268  0.7774  0.8793
```

N = 1975, Events: 1006, Censored: 969

1) Total time at risk: 9192.058

Log-likelihood = -2906.248, df = 4

AIC = 5820.497

*New treatment increases time
to progression (AFT model)*

Shape parameter: whether the hazard is increasing (>1), decreasing (<1), or constant (=1)

Scale: whether the hazard is lower/higher at given time point

Parameter estimation – Multi-state model

Stable -> Death

```
[[2]]
Call:
flexsurvreg(formula = Surv(time, status) ~ strategy_name + female,
  data = data, subset = (transition_id == i), dist = "weibull")

Estimates:
            data  mean    est      L95%     U95%      se   exp(est)      L95%     U95%
shape          NA  2.482459  2.303776  2.675000  0.094614      NA        NA        NA
scale          NA 10.406898  9.612760 11.266641  0.421473      NA        NA        NA
strategy_nameNew 0.528101  0.241482  0.157695  0.325268  0.042749  1.273134  1.170810  1.384402
female         0.494684 -0.083337 -0.167300  0.000626  0.042839  0.920041  0.845945  1.000626
```

N = 1975, Events: 358, Censored: 1617

Total time at risk: 9192.058

Log-likelihood = -1333.968, df = 4

AIC = 2675.936

Shape parameter: whether the hazard is increasing, decreasing, or constant

Scale: whether the hazard is lower/higher at given time point

Parameter estimation – Multi-state model

Progression > Death

```
[[3]]  
Call:  
flexsurvreg(formula = Surv(time, status) ~ strategy_name + female,  
            data = data, subset = (transition_id == i), dist = "weibull")  
  
Estimates:  
          data mean    est      L95%     U95%      se exp(est)    L95%     U95%  
shape        NA  3.48340  3.25461  3.72826  0.12074      NA      NA      NA  
scale        NA  8.96768  8.55835  9.39658  0.21376      NA      NA      NA  
strategy_nameNew 0.50398  0.00922 -0.04311  0.06155  0.02670  1.00926  0.95780  1.06348  
female       0.52386 -0.11650 -0.16914 -0.06385  0.02686  0.89003  0.84439  0.93815
```

N = 1006, Events: 468, Censored: 538

Total time at risk: 5479.46

Log-likelihood = -1237.573, df = 4

AIC = 2483.147

Shape parameter: whether the hazard is increasing, decreasing, or constant

Scale: whether the hazard is lower/higher at given time point

Parameters – Utility

```
utility_tb1 <- stateval_tb1(
  data.table(state_id = states$state_id,
            mean = c(.8, .6),
            se = c(0.02, .05)
  ),
  dist = "beta")
```

```
state_id mean   se
1:       1  0.8 0.02
2:       2  0.6 0.05
```

Parameters – Medical cost

```
medcost_tb1 <- stateval_tb1(  
  data.table(state_id = states$state_id,  
            mean = c(2000, 9500),  
            se = c(2000, 9500)  
,  
  dist = "gamma")
```

```
state_id mean   se  
1:          1 2000 2000  
2:          2 9500 9500
```

Parameters – Drug cost

```
n_times <- 2

drugcost_tbl <- stateval_tbl(
  data.table(
    strategy_id = rep(strategies$strategy_id, each = n_states * n_times),
    state_id = rep(rep(states$state_id, each = n_strategies), n_times),
    time_start = rep(c(0, 3/12), n_states * n_strategies),
    est = c(rep(2000, 4), # Costs are always the same with SOC
           12000, 12000, 12000, 10000 # Costs with New drop after 3 months in progression state
    )
  ),
  dist = "fixed")
```

When using an IPS, "state values" (like transition rates) can depend on time in an intermediate health state

We illustrate by assuming that costs for the new treatment are \$12,000 for the first 3 months in the progression state and then \$10,000 thereafter

(Would not be possible in a cohort model without creating tunnel states)

	strategy_id	state_id	time_id	time_start	time_stop	est
1:		1	1	0.00	0.25	2000
2:		1	1	0.25	Inf	2000
3:		1	2	0.00	0.25	2000
4:		1	2	0.25	Inf	2000
5:		2	1	0.00	0.25	12000
6:		2	1	0.25	Inf	12000
7:		2	2	0.00	0.25	12000
8:		2	2	0.25	Inf	10000
						104

Simulation – Construct the model

Disease model

- The transition model is constructed as a function of the fitted multi-state model and *input data* (treatment strategy and patients)

```
transmod_data <- expand(hesim_dat,
                         by = c("strategies", "patients"))
```

	strategy_id	patient_id	strategy_name	age	female
1:	1	1	SOC	42.71774	0
2:	1	2	SOC	48.86723	0
3:	1	3	SOC	40.27539	1
4:	1	4	SOC	46.50052	1
5:	1	5	SOC	47.17538	1
6:	1	6	SOC	53.21776	0

```
transmod <- create_IndivCtstmTrans(wei_fits, transmod_data,
                                    trans_mat = tmat, n = 500,
                                    clock = "reset",
                                    start_age = patients$age)
```

Simulation – Construct the model

Utility and cost models

```
# Utility
utilitymod <- create_StateVals(utility_tbl, n = 500,
                                 hesim_data = hesim_dat)

# Costs
drugcostmod <- create_StateVals(drugcost_tbl, n = 500,
                                   time_reset = TRUE, ←
                                   hesim_data = hesim_dat) So that costs depend on  
time in intermediate state

medcostmod <- create_StateVals(medcost_tbl, n = 500,
                                 hesim_data = hesim_dat)

costmods <- list(Drug = drugcostmod,
                  Medical = medcostmod)
```

Simulation – Construct the model

Combining the disease progression, cost, and utility models

```
econmod <- IndivCtstm$new(trans_model = transmod,  
                           utility_model = utilitymod,  
                           cost_models = costmods)
```

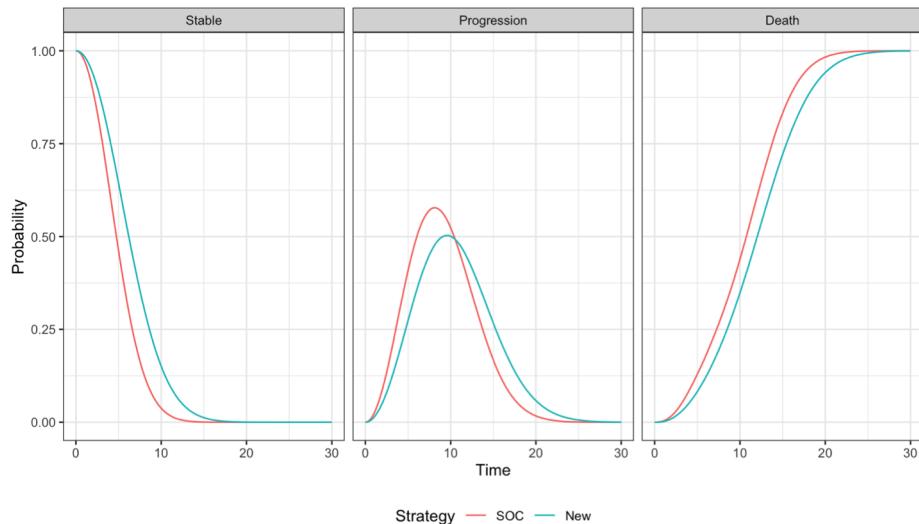
Simulation - Simulating outcomes

Disease progression

```
econmod$sim_disease(max_age = 100)  
head(econmod$disprog_)
```

	sample	strategy_id	patient_id	grp_id	from	to	final	time_start	time_stop
1:	1		1	1	1	2	0	0.000000	1.420130
2:	1		1	1	1	2	3	1	1.420130
3:	1		1	2	1	1	2	0	0.000000
4:	1		1	2	1	2	3	1	5.711780
5:	1		1	3	1	1	2	0	0.000000
6:	1		1	3	1	2	3	1	5.249609
									9.059670

```
econmod$sim_stateprobs(t = seq(0, 30 , 1/12))  
autoplot(econmod$stateprobs_, labels = labs)
```



Simulation - Simulating outcomes

QALYs and costs

```
econmod$sim_qalys(dr = c(0,.03))
```

```
sample strategy_id grp_id state_id dr      qalys      lys
1:     1             1       1           1 0 3.795199 5.009791
2:     1             1       1           2 0 3.433499 5.368864
3:     1             2       1           1 0 4.898627 6.466354
4:     1             2       1           2 0 3.491527 5.459601
5:     2             1       1           1 0 3.882274 4.793997
6:     2             1       1           2 0 3.220405 5.746082
```

```
econmod$sim_costs(dr = 0.03)
```

```
sample strategy_id grp_id state_id dr category      costs
1:     1             1       1           1 0.03    Drug  9124.834
2:     1             1       1           2 0.03    Drug  8261.846
3:     1             2       1           1 0.03    Drug  69025.893
4:     1             2       1           2 0.03    Drug  40511.631
5:     2             1       1           1 0.03    Drug  8778.825
6:     2             1       1           2 0.03    Drug  8863.685
```

Cost-effectiveness analysis

```
ce_sim <- econmod$summarize()

cea_pw_out <- cea_pw(ce_sim, comparator = 1,
                      dr_qalys = .03, dr_costs = .03)

format(icer(cea_pw_out, labels = 1abs))
```

	Outcome	New
1:	Incremental QALYs	0.86 (0.56, 1.17)
2:	Incremental costs	92,981 (81,657, 101,697)
3:	Incremental NMB	-50,160 (-61,751, -38,254)
4:	ICER	108,571

Steps with `hesim`

1. Model set-up

- Specify the treatment strategies, target population(s), and model structure
 - `hesim_data()`

2. Parameters

- Estimate or define the parameters of the economic model
 - `flexsurvreg_list()`, `stateval_tb1()`

3. Simulation

a. Construction of model

- Create an economic model—consisting of separate statistical models for disease progression, costs, and utilities—that simulate outcomes as a function of *input data* (derived from Step 1) and *parameters* (from Step 2)
- `create_IndivCtstmTrans()`, `create_Statevals()`, `IndivCtstm$new()`

b. Simulation of outcomes

- Simulate outcomes (disease progression, costs, and quality-adjusted life-years (QALYs)) using the model constructed in Step 3
- `$sim_disease()`, `$sim_stateprobs()`, `$sim_qalys()`, `$sim_costs()`

4. Cost-effectiveness analysis

Complete R script

The screenshot shows an RStudio interface with a script file titled "04-mstate.R". The code is a complete R script for state transition modeling, using packages like rcea, hesim, data.table, ggplot2, and flexsurv. It includes sections for library imports, setting random seeds, defining transition matrices, generating patient data, and creating state and strategy data frames. The code is annotated with comments and uses various R functions like rbind, rnorm, and rbinom.

```
## ---- Overview -----
## @knitr R-setup
library("rcea")
library("hesim")
library("data.table")
library("ggplot2")
library("flexsurv")
theme_set(theme_bw())

set.seed(101) # Make random number generation reproducible

## ---- Model setup -----
## @knitr tmatrix
tmatrix <- rbind(
  c(NA, 1, 2),
  c(NA, NA, 3),
  c(NA, NA, NA)
)
colnames(tmatrix) <- rownames(tmatrix) <- c("Stable", "Progression", "Dead")
print(tmatrix)

## @knitr hesim_data
n_patients <- 1000
patients <- data.table(
  patient_id = 1:n_patients,
  age = rnorm(n_patients, mean = 45, sd = 7),
  female = rbinom(n_patients, size = 1, prob = .51)
)

states <- data.table(
  state_id = c(1, 2),
  state_name = c("Stable", "Progression") # Non-death health states
)
n_states <- nrow(states)

strategies <- data.frame(
  strategy_id = 1:2,
  strategy_name = c("SOC", "New")
)
n_strategies <- nrow(strategies)

hesim_dat <- hesim_data(
  strategies = strategies,
  patients = patients,
  states = states
)
```

Tutorial

rcea **0.1.2**

Reference

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Slides



Semi-Markov Multi-state Model

2021-07-26

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Overview

RATHERESTIMATION

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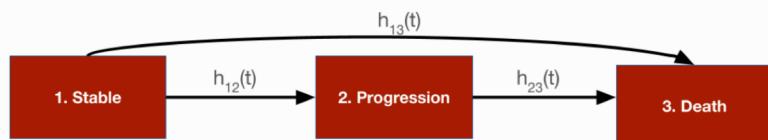
Cost-effectiveness analysis

<https://hesim-dev.github.io/rcea/articles/04-mstate.html>

Overview

In this tutorial we use a continuous time state transition model (CTSTM) and relax many of the assumptions made in cohort discrete time state transition models (DTSTMs). First, since the model is in continuous time we do not require model cycles. Second, we estimate the parameters of the health state transitions using a multi-state model so that the simulation model is completely integrated with an underlying statistical model. Third, we use individual patient simulation (IPS) to simulate a semi-Markov model, meaning that (unlike in a Markov model) transitions cannot depend on prior history.

To illustrate, we simplify the sick-sicker model so that it only contains three health states and modify the states—*Stable*, *Progression*, and *Dead*—to mimic an oncology application where patients transition from stable disease to progression to death. There are three transitions: (1) *Stable* to *Progression*, (2) *Stable* to *Dead*, and (3) *Progression* to *Dead*.



The following packages and settings will be used for the analysis. Note that while individual-level simulations can be computationally intensive, they run very quickly in `hesim` because they are implemented fully in C++ under the hood. You can learn more by looking at the `hesim` multi-state modeling vignette.

Exercise 4: Semi-Markov multi-state model

- *Modify R-script “04-mstate.R”*
 - *Simulate homogeneous patient cohort*
 - *Use a generalized gamma distribution for transitions (hint: type ?flexsurvreg into R and look at the options for the dist argument)*
- *Run modified script*

Partitioned survival model

Steps with `hesim`

1. Model set-up

- Specify the treatment strategies, target population(s), and states
 - `hesim_data()`

2. Parameters

- Estimate or define the parameters of the economic model
 - `flexsurvreg_list()`, `stateval_tb1()`

3. Simulation

a. Construction of model

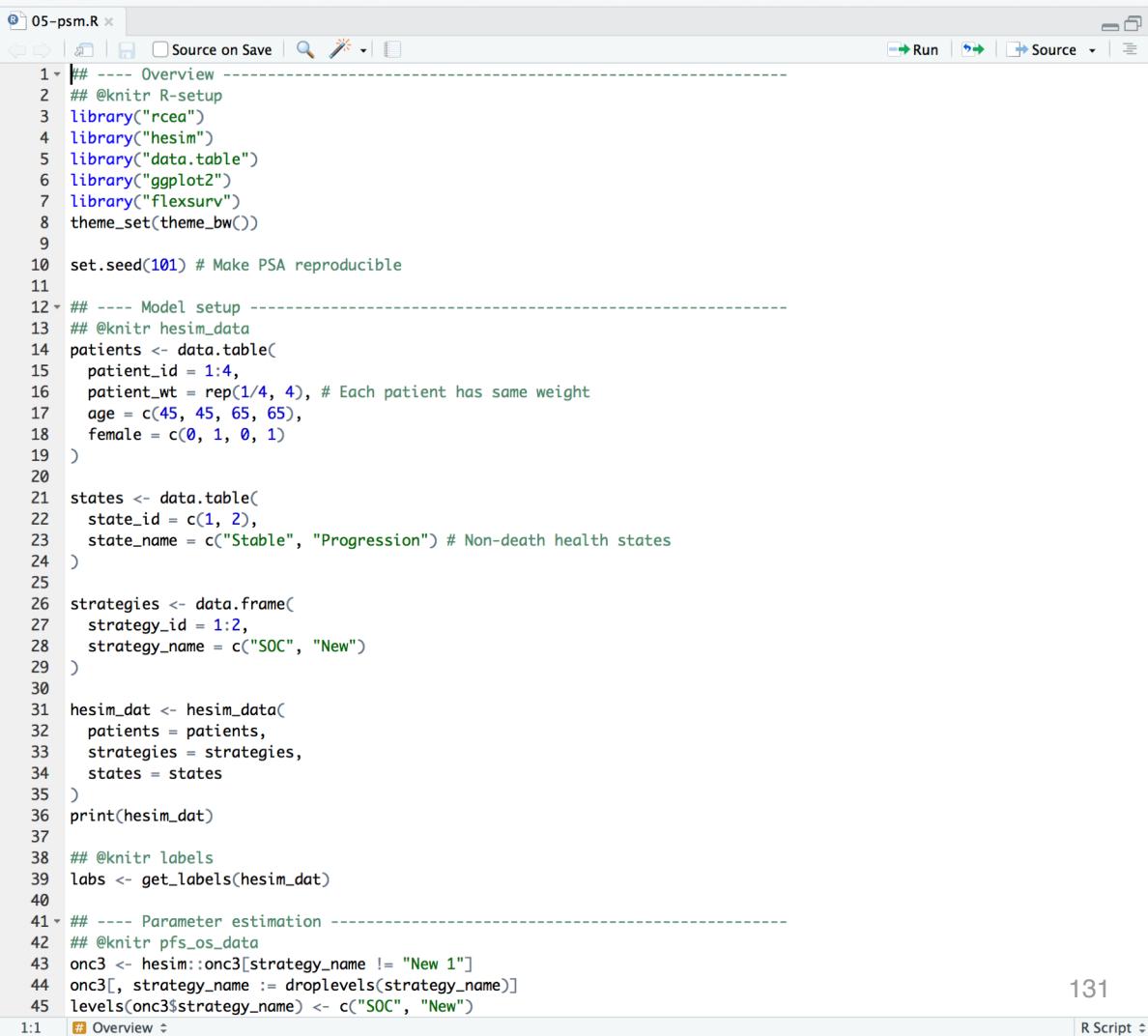
- Create an economic model—consisting of separate statistical models for disease progression (survival models), costs, and utilities—that simulate outcomes as a function of *input data* (derived from Step 1) and *parameters* (from Step 2)
- `create_PsmCurves()`, `create_Statevals()`, `Psm$new()`

b. Simulation of outcomes

- Simulate outcomes using the model constructed in Step 3
- `$sim_survival()`, `$sim_stateprobs()`, `$sim_qalys()`, `$sim_costs()`

4. Cost-effectiveness analysis

Complete R script



```
## ---- Overview ----
## @knitr R-setup
library("rcdd")
library("hesim")
library("data.table")
library("ggplot2")
library("flexsurv")
theme_set(theme_bw())
set.seed(101) # Make PSA reproducible
## ---- Model setup ----
## @knitr hesim_data
patients <- data.table(
  patient_id = 1:4,
  patient_wt = rep(1/4, 4), # Each patient has same weight
  age = c(45, 45, 65, 65),
  female = c(0, 1, 0, 1)
)
states <- data.table(
  state_id = c(1, 2),
  state_name = c("Stable", "Progression") # Non-death health states
)
strategies <- data.frame(
  strategy_id = 1:2,
  strategy_name = c("SOC", "New")
)
hesim_dat <- hesim_data(
  patients = patients,
  strategies = strategies,
  states = states
)
print(hesim_dat)
## @knitr labels
labs <- get_labels(hesim_dat)
## ---- Parameter estimation ----
## @knitr pfs_os_data
onc3 <- hesim::onc3[strategy_name != "New 1"]
onc3[, strategy_name := droplevels(strategy_name)]
levels(onc3$strategy_name) <- c("SOC", "New")
```

Tutorial

rcea **0.1.2**

Reference

Tutorials ▾

Slides



Partitioned Survival Model

2021-07-26

<https://hesim-dev.github.io/rcea/articles/05-psm.html>

Overview

While multi-state models can be used to estimate the parameters of a state transition model (STM) in a very flexible manner, data availability can make it difficult (or infeasible) to fit such a model. This is often the case when an evidence synthesis model based on summary level data is used to parameterize the STM. For example, in oncology, published articles of clinical trials often provide survival curves of progression-free survival (PFS) and overall survival (OS), but do not release information on time to event (and censoring) for each transition. In this setting partitioned survival analysis may consequently be a simpler approach.

We will use the same packages as in the ["Semi-Markov Multi-state Model"](#) tutorial.

```
library("rcea")
library("hesim")
library("data.table")
library("ggplot2")
library("flexsurv")
theme_set(theme_bw())

set.seed(101) # Make PSA reproducible
```

Contents

Overview

Theory

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Cost-effectiveness analysis

Theory

An 3-state partitioned survival model (PSM) simulates the probability that a patient is in each of 3 distinct health states at a given point of time when treated by a particular therapy. State membership is estimated from 2 survival curves (e.g., PFS and OS) using an "area under the

Cost-effectiveness analysis

Cost-effectiveness analysis

- The optimal treatment strategy is the one that maximizes the expected net monetary benefit (NMB)

$$NMB(j, \theta) = e_j(\theta) \cdot k - c_j(\theta)$$

Effectiveness
(e.g., QALYs) Willingness
to pay Costs

j indexes a treatment strategy
θ indexes a parameter set

$$j^* = \operatorname{argmax}_j E_\theta[NMB(j, \theta)]$$

*j** is treatment with highest NMB
averaged across all sets of *θ*

- Can also assess optimal strategy using incremental cost-effectiveness ratio (ICER). Treatment 1 is preferred to treatment 0 if¹

$$k > \frac{E_\theta[c_1(\theta) - c_0(\theta)]}{E_\theta[e_1(\theta) - e_0(\theta)]} = ICER$$

¹Only true if both incremental costs (numerator) and incremental effectiveness (denominator) are both positive. Treatment 1 dominates treatment 0 if it is more effective and less costly. Treatment 1 is dominated by treatment 0 if it is less effective and more costly. Treatment 1 is preferred to treatment 0 if it is less costly and less effective when $k < ICER$.

Value of perfect information

- The expected value of perfect information (EVPI) combines the probability of being most effective with the *magnitude* of the expected NMB
- Intuitively, EVPI is the amount that a decision maker would be willing to pay to collect additional data and completely eliminate uncertainty
- Mathematically, the EVPI is defined as the difference between the maximum expected NMB given perfect information and the maximum expected NMB given current information

$$EVPI = E_{\theta} [\max_j NMB(j, \theta)] - \max_j E_{\theta} [NMB(j, \theta)]$$

NMB for optimal treatment at each random draw of the parameters

NMB of treatment that is optimal when averaged across all parameter draws

Economic models with **hesim**

1. Model set-up
2. Parameters
3. Simulation
 - a. Construction of model
 - b. Simulation of outcomes
4. Cost-effectiveness analysis

Cost-effectiveness analysis with `hesim`

- `hesim` can be used to perform CEA and summarize decision uncertainty
 - Other R packages such as BCEA, dampack, SAVI, and EVSI could also be considered, especially for more advanced value of information analysis
- Implementation via the `cea()` and `cea_pw()` functions
 - `cea()` summarizes results by taking into account each treatment strategy in the analysis
 - `cea_pw()` summarizes “pairwise” results in which each treatment is compared to a comparator
- Both are “generic” functions that work with (i) data frame like objects or (ii) “ce” objects

Cost-effectiveness analysis with hesim

```
markov_hesim_ce <- readRDS("markov-cohort-hesim-ce_sim.rds")
```

```
markov_hesim_ce
```

```
$costs
  category dr sample strategy_id      costs grp_id
1: treatment 0.03     1           1 52301.47     1
2: treatment 0.03     1           2 323892.01     1
3: treatment 0.03     2           1 49767.92     1
4: treatment 0.03     2           2 322401.36     1
5: treatment 0.03     3           1 52661.20     1
---
```

```
5996:   total 0.03    998           2 434677.65     1
5997:   total 0.03    999           1 165176.48     1
5998:   total 0.03    999           2 446988.07     1
5999:   total 0.03   1000           1 79522.29     1
6000:   total 0.03   1000           2 360039.77     1
```

```
$qalys
  dr sample strategy_id      qalys grp_id
1: 0.03     1           1 20.90995     1
2: 0.03     1           2 22.24340     1
3: 0.03     2           1 20.77562     1
4: 0.03     2           2 23.51086     1
5: 0.03     3           1 20.89219     1
---
```

```
1996: 0.03    998           2 24.15825     1
1997: 0.03    999           1 20.88165     1
1998: 0.03    999           2 23.63537     1
1999: 0.03   1000           1 20.77720     1
2000: 0.03   1000           2 23.02164     1
```

```
wtp <- seq(0, 250000, 500) #willingness to pay per QALY

cea_pw_out <- cea_pw(markov_hesim_ce, comparator = "SOC",
                       sample = "sample", strategy = "strategy",
                       e = "dqlalys", c = "dcosts",
                       k = wtp
)

cea_out <- cea(markov_hesim_ce,
                 sample = "sample", strategy = "strategy",
                 e = "dqlalys", c = "dcosts",
                 k = wtp
)
```

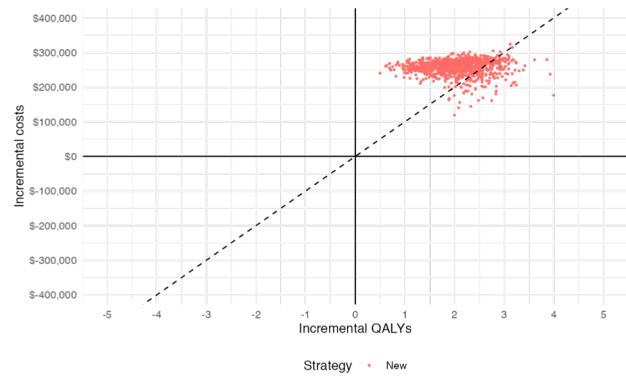
Incremental cost-effectiveness ratio with **hesim**

```
icer(cea_pw_out, wtp = 50000) %>%  
  format()
```

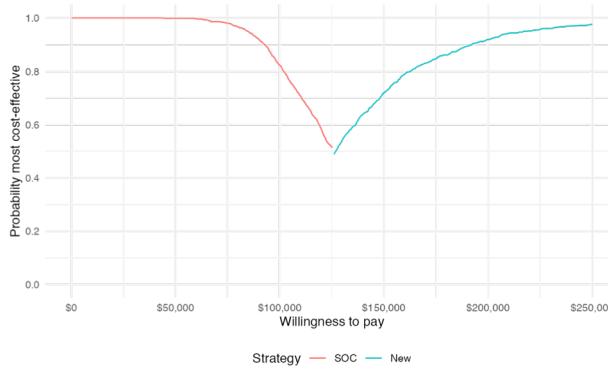
	Outcome	New
1: Incremental QALYs		2.09 (1.04, 3.14)
2: Incremental costs	261,594	(212,238, 291,654)
3: Incremental NMB	-156,981	(-215,676, -83,866)
4: ICER		125,029

Representing decision uncertainty with hesim

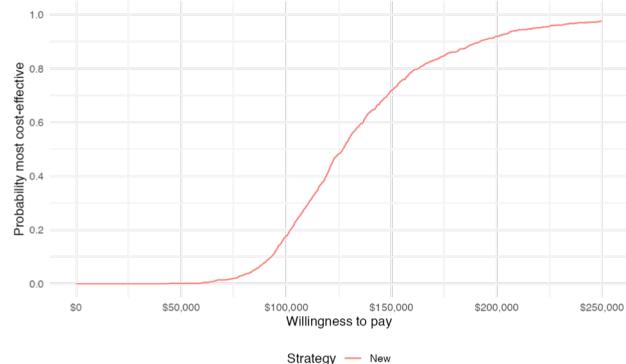
plot_ceplane(cea_pw_out)



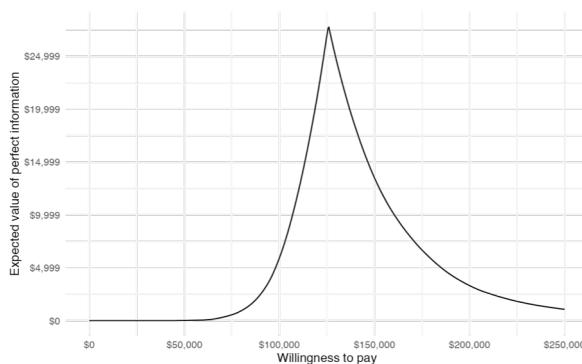
plot_ceaf(cea_out)



plot_ceac(cea_pw_out)



plot_evpri(cea_out)



Complete R script

```
06-cea.R x
Source on Save | Run | Source | Overview

1 ## ---- Overview -----
2 ## @knitr R-setup
3 library("hesim")
4 library("ggplot2")
5 library("magrittr")
6 theme_set(theme_minimal()) # Set ggplot2 theme
7
8 ## ---- Application -----
9 ## @knitr load-ce
10 markov_ce <- readRDS("markov-cohort-ce_sim.rds")
11 markov_ce
12
13 ## @knitr load-hesim-ce
14 markov_hesim_ce <- readRDS("markov-cohort-hesim-ce_sim.rds")
15 hesim_dat <- readRDS("markov-cohort-hesim_data.rds")
16 markov_hesim_ce
17
18 ## @knitr conduct-cea
19 wtp <- seq(0, 250000, 500) # Willingness to pay per QALY
20 cea_pw_out <- cea_pw(markov_hesim_ce,
21                         comparator = 1, # Comparator is SOC (ID = 1)
22                         dr_qalys = 0.03, dr_costs = 0.03,
23                         wtp)
24 cea_out <- cea(markov_hesim_ce,
25                  dr_qalys = 0.03, dr_costs = 0.03,
26                  k = wtp)
27
28 ## @knitr conduct-cea-default
29 cea_pw_out2 <- cea_pw(markov_ce, comparator = "SOC",
30                        k = wtp,
31                        sample = "sample", strategy = "strategy",
32                        e = "dqalys", c = "dcosts")
33 cea_pw_out$summary
34 cea_pw_out2$summary
35
36 ## ---- Incremental cost-effectiveness ratio -----
37 ## @knitr icer
38 labs <- get_labels(hesim_dat)
39 icer(cea_pw_out, wtp = 50000, labels = labs) %>%
40   format()
41
42 ## ---- Cost-effectiveness plane -----
43 ## @knitr ceplane-plot
44 plot_ceplane(cea_pw_out, k = 100000, labels = labs)
45
```

Tutorial

rcea **0.1.2**

Reference

Tutorials ▾

Slides



Cost-effectiveness Analysis

2021-07-26

Contents

Overview

<https://hesim-dev.github.io/rcea/articles/06-cea.html>

CUSTOM PUBLISHING WITH GITHUB

Overview

The prior tutorials have focused on constructing economic models to simulate disease progression, costs, and quality-adjusted life-years (QALYs). While incremental cost-effectiveness ratios (ICERs) have been computed and probabilistic sensitivity analysis (PSA) has been employed, we have not yet formalized cost-effectiveness analysis (CEA) or represented decision uncertainty.

In this analysis we will perform a CEA given the output of model from the “[Semi-Markov Multi-state Model](#)” tutorial. We will use the CEA functions from `hesim` to summarize decision uncertainty and `ggplot2` for visualization. The CEA will be performed for a single target population, but you can review the `hesim` tutorial on [CEA](#) and the references therein for an example of CEA in the context of multiple subgroups.

```
library("hesim")
library("ggplot2")
library("magrittr")
theme_set(theme_minimal()) # Set ggplot2 theme
```

Theory

CEA is based on estimating the net monetary benefit (NMB). For a given parameter set θ , the NMB with treatment j is computed as the difference between the monetized health gains from an intervention less costs, or,

$$NMB(j, \theta) = e_j(\theta) \cdot k - c_j(\theta),$$

where e_j and c_j are measures of health outcomes (e.g. QALYs) and costs using treatment j respectively, and k is a decision makers

Summary

So why R for CE modeling?

- One platform to do everything
 - parameter estimation
 - simulation
- More complex analysis and individual patient simulation
- Your problems are rarely unique
 - But even if they are, R facilitates development of custom models
- Easier to share and review
- Reproducible

Cost-effectiveness analysis with R

- BCEA
- heemod
- hesim
- ...

Why **hesim**?

- Focus on setting up and interpreting a model rather than implementation
 - Burdensome programming tasks have already been implemented and optimized
- Flexible enough to cover many problems
 - May need to write custom code for very complex or unique problems (beyond scope of course)
- Very fast!
 - IPS in **hesim** with 100 PSA iterations ran in .44 seconds; equivalent simulation in **mstate** package took 34 minutes (see [here](#))
 - Cohort model in **hesim** with 1,000 PSA iterations ran in 1 second (IPS in 9 seconds), while equivalent cohort model in **heemod** took 85 seconds (see [here](#))

User interfaces with R Shiny

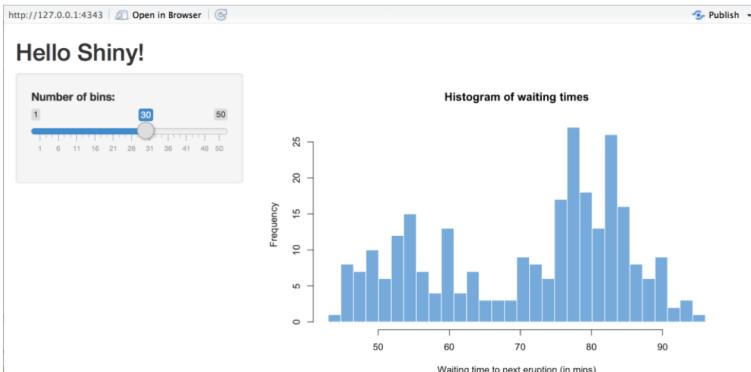
LESSON 1

Welcome to Shiny

Shiny is an R package that makes it easy to build interactive web applications (apps) straight from R. This lesson will get you started building Shiny apps right away.

```
install.packages("shiny")
```

Examples



<https://shiny.rstudio.com/>

Making Markov Models Shiny: A Tutorial

Robert Smith and Paul Schneider, SchARR, University of Sheffield

Sick Sicker Model in Shiny

Treatment Cost
200

PSA runs
1000

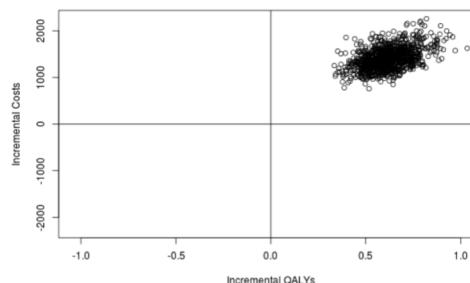
initial age
10 25 80

Run / update model

Results Table

Option	QALYs	Costs	Inc.QALYs	Inc.Costs	ICER
Treatment	18.59	100441.67	0.62	1406.24	2324.54
No Treatment	17.97	99035.43	NA	NA	NA

Cost-effectiveness Plane



https://r-hta.org/tutorial/markov_models_shiny/

[Restore defaults](#)

Value of a QALY ⓘ \$ 150,000

[Save](#)[Run simulation](#)

Treatment sequences ⓘ

[Show survival curves](#)

Line of therapy

Treatment regimen

Sequence number

I II III IV V

1L

gefitinib

erlotinib

afatinib

dacomitinib

osimertinib

T790M Status + - + - + -

2L

gefitinib

erlotinib

afatinib

dacomitinib

osimertinib

pbdc

pbdc + bevacizumab

pbdc

pbdc + nivolumab

pbdc + pembrolizumab

pbdc + atezolizumab

pbdc + bevacizumab

pbdc + bevacizumab + nivolumab

Cost-effectiveness plane ⓘ

Sequence-comparator

I

II

III

IV

Expected outcomes

ICER

CE PLANE

CEAC

● Sequence II ● Sequence III ● Sequence IV — Value of a QALY

\$ 1,800,000

\$ 1,000,000

\$ 0

-\$ 1,000,000

-\$ 1,800,000

Incremental costs

-8.00 -6.00 -3.00 0.0 3.00 6.00 8.00

Incremental QALYs

Expected value of perfect information

 Restore defaults

Value of a QALY ⓘ \$ 150,000

 Save Run simulationR Code 

```
22 pats <- create_patients(n = 100)

23

24

25 txseq1 <- txseq(
26   first = c("gefitinib"),
27   second = c("osimertinib", "PBDC"),
28   second_plus = c("PBDC + atezolizumab", "PBDC + atezolizumab")
29 )

30

31 txseq2 <- txseq(
32   first = c("erlotinib"),
33   second = c("osimertinib", "PBDC"),
34   second_plus = c("PBDC + atezolizumab", "PBDC + atezolizumab")
35 )

36

37

38 txseq3 <- txseq(
39   first = c("afatinib"),
40   second = c("osimertinib", "PBDC"),
41   second_plus = c("PBDC + atezolizumab", "PBDC + atezolizumab")
42 )

43

44

45 txseq4 <- txseq(
46   first = c("osimertinib"),
47   second = c("PBDC", "PBDC"),
48   second_plus = c("PBDC + atezolizumab", "PBDC + atezolizumab")
```

pbdc + bevacizumab

pbdc + bevacizumab + nivolumab

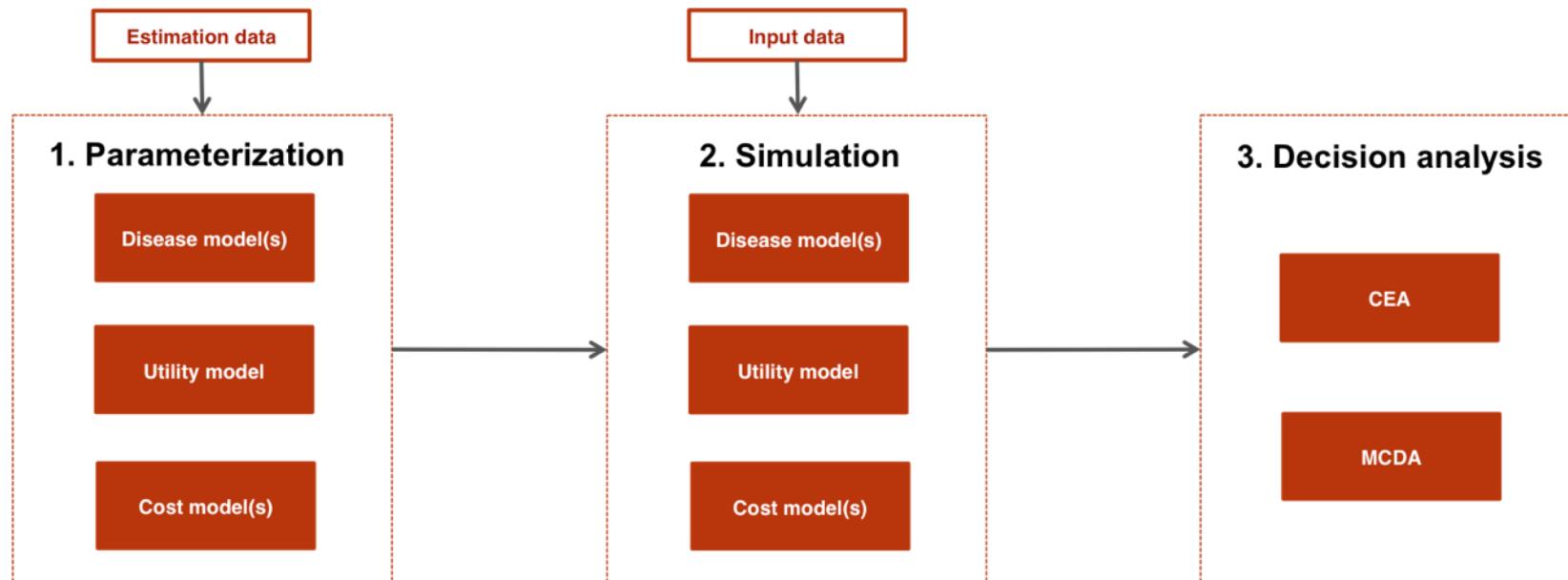


Thank you

devin.incerti@gmail.com

jeroen.jansen@ucsf.edu

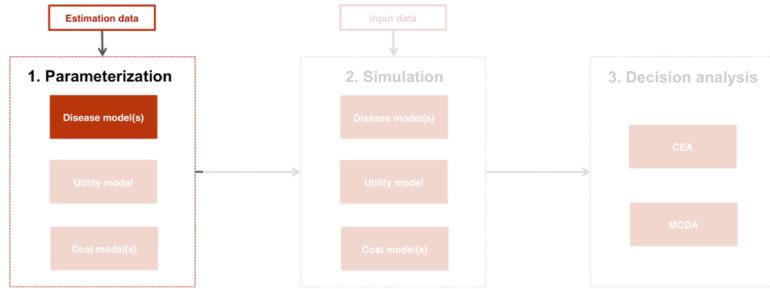
Appendix – Building a model with **hesim**



hesim overview

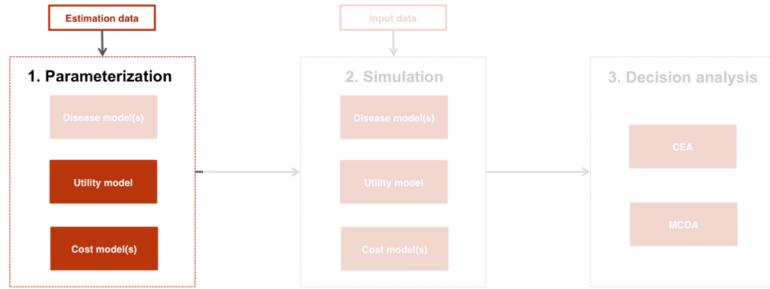
Economic model	<i>Object</i>	<i>Function</i>
Cohort discrete time state transition model (cDTSTM)	<code>CohortDtstm</code>	<code>create_CohortDtstm()</code> <code>CohortDtstm\$new()</code>
Individual-level continuous time state transition model (iCTSTM)	<code>IndivCtstm</code>	<code>IndivCtstm\$new()</code>
N-state partitioned survival model (PSM)	<code>Psm</code>	<code>Psm\$new()</code>

hesim - Parameterization



		Disease progression		
		<i>Statistical model</i>	<i>Parameter Object</i>	<i>Model fit Function</i>
cDTSTM	<code>CohortDtstm</code>	Custom	<code>tparams_transprobs</code>	<code>define_model()</code>
		Multinomial logistic regression	<code>params_mlogit_list</code>	<code>multinom_list()</code>
iCTSTM	<code>IndivCtstm</code>	Multi-state model (joint likelihood)	<code>params_surv</code>	<code>flexsurv::flexsurvreg()</code>
		Multi-state model (transition-specific)	<code>params_surv_list</code>	<code>flexsurvreg_list()</code>
PSM	<code>Psm</code>	Independent survival models	<code>params_surv_list</code>	<code>flexsurvreg_list()</code>

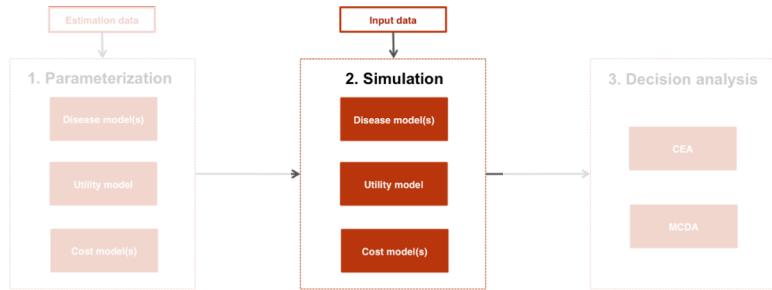
hesim - Parameterization



Cost and utility		
<i>Statistical model</i>	<i>Parameter Object</i>	<i>Model fit Function</i>
Predicted means	<code>tparams_mean</code>	<code>stateval_tb1()</code>
	<code>tparams_mean</code>	<code>define_model()</code>
Linear model	<code>params_lm</code>	<code>Stats::lm()</code>

hesim - Simulation

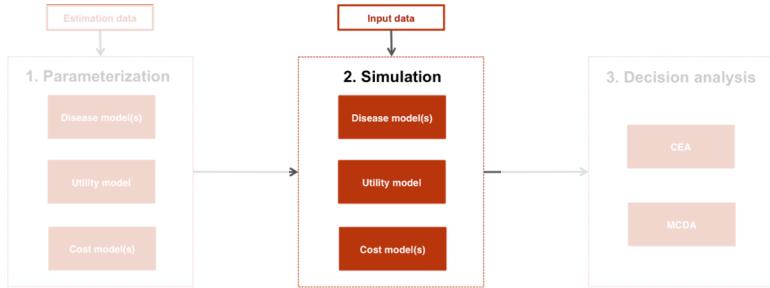
Constructing an economic model



Economic model		Disease model	Utility model	Cost model(s)
cDTSTM	CohortDtstm	CohortDtstmTrans create_CohortDtstmTrans()	Statevals create_Statevals()	Statevals create_Statevals()
iCTSTM	IndivCtstm	IndivCtstmTrans create_IndivCtstmTrans()	Statevals create_Statevals()	Statevals create_Statevals()
PSM	Psm	PsmCurves create_PsmCurves()	Statevals create_Statevals()	Statevals create_Statevals()

hesim - Simulation

Simulating outcomes



Economic model		Disease progression	QALYs	Costs
cDTSTM	CohortDtstm	\$sim_stateprobs()	\$sim_qalys()	\$sim_costs()
iCTSTM	IndivCtstm	\$sim_disease() \$sim_stateprobs()	\$sim_qalys()	\$sim_costs()
PSM	Psm	\$sim_survival() \$sim_stateprobs()	\$sim_qalys()	\$sim_costs()